



Research Article

# Beetles (Coleoptera) in deciduous dead wood tree species trunks in Lithuania

Aistė Lekoveckaitė<sup>‡</sup>, Virginija Podénienė<sup>‡</sup>, Romas Ferenca<sup>§</sup>

<sup>‡</sup> Vilnius University, Life Sciences Center, Vilnius, Lithuania

<sup>§</sup> Kaunas T. Ivanauskas Zoological Museum, Kaunas, Lithuania

Corresponding author: Aistė Lekoveckaitė ([aiste.lekoveckaite@gmc.vu.lt](mailto:aiste.lekoveckaite@gmc.vu.lt))

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## Abstract

We present a list of beetles that emerged from wind-felled tree trunks of several tree species, including European ash (*Fraxinus excelsior*), aspen (*Populus tremula*), common oak (*Quercus robur*), birch (*Betula* sp.), small-leaved linden (*Tilia cordata*) and black alder (*Alnus glutinosa*). Four hundred and ninety species and 60 families of beetles were collected using trunk-emergence type traps. We found 440 beetle species that had previously been recorded from dead wood; the remaining 50 were newly discovered and all were considered as not directly associated with dead wood. Common oak trunks had the highest diversity of beetles, with approximately 42% of the identified beetle species found in our research. Of all the beetle species identified in the study, about half are saproxylic, while the remaining are considered as not having direct association with dead wood. The results of the study emphasise the importance of dead wood in maintaining beetle diversity in Lithuanian forests. This study provides a valuable baseline for future research on beetles in dead wood in Lithuania and may help to provide information for conservation efforts to protect these important habitats.

## Keywords

saproxylic, trunk-emergence traps, wind-felled trees

## Introduction

Forest land occupies 33.8% of Lithuania (Government of the Republic of Lithuania 2022). Lithuanian forests belong to the European hemi-boreal mixed broadleaved-coniferous forest type and around 44% of the country's forests consist of deciduous tree species (Varnagirytė-Kabašinskienė et al. 2019). Amongst them, *Betula* sp. occupies 22.2% of the forest area, *Populus tremula* stands occupy 4.6%, *Alnus glutinosa* and *Alnus incana* occupy 7.6% and 5.9%, respectively, *Quercus robur* and *Fraxinus excelsior* occupy 2.2% and 0.9%, respectively, while areas covered by other deciduous tree species are insignificant (1.1%) (Varnagirytė-Kabašinskienė et al. 2019). According to the recent state accounting of Lithuanian forests, which summarises information about forest resources, their quality, natural and economic condition, the trends in the species composition of stands have changed little over the past few years (Government of the Republic of Lithuania 2022). However, areas of conifer, black alder and oak stands are increasing, while areas covered by birch, aspen, white alder and ash are shrinking (Government of the Republic of Lithuania 2022).

In a healthy forest, wood comes in many forms, including living trees, stumps, snags, logs and branches (Paletto and Tosi 2010). These forms of wood host a variety of species, mainly including *fungi* and insects (Stokland et al. 2012, Jacobsen et al. 2015). One of the most important group of insects are saproxylic beetles, which depend on dead or dying wood for some part of their life cycle (Speight 1989). Beetles play an important role in decomposing and recycling dead wood. In fact, more than half of forest-dwelling beetles may be saproxylic (Bouget et al. 2008). Beetles can be grouped, based on their lifestyle – obligate or facultative saproxyls (e.g. Maňák and Schlaghamerský (2009), Milberg et al. (2014), Milberg et al. (2016)), some are flexible and can live on a variety of dead tree species, while others are specific to one type of host tree (Jonsell et al. 1998, Abrahamsson et al. 2009, Toivanen and Kotiaho 2010, Milberg et al. 2014).

Numerous studies have been conducted to investigate various aspects of beetle assemblages, including their relationships to environmental conditions and associations with different tree species (Horák 2011, Macagno et al. 2015, Muñoz-López et al. 2016, Procházka and Schlaghamerský 2019, Zuo et al. 2021, Edelmann et al. 2022). Tree species identity has been identified as one of the most significant drivers determining the community composition of beetles (Müller et al. 2020). While the number of species showing strict host-specificity is relatively low (Jonsson et al. 2005), beetle communities, in general, are dependent on tree species and this dependence decreases significantly as decay progresses (Wende et al. 2017, Zuo et al. 2021). According to research in north Europe (Jonsson et al. 2005), out of nearly 7000 wood-living species in Sweden, only around 130 have been found to exclusively inhabit a single tree species. Additionally, birch, oak and aspen are amongst the most species-rich deciduous trees in the country. Taking south-eastern Sweden as an example, 19 out of 171 beetle species had a significant association with common oak, six with Norway maple, two with European ash and five with small-leaved linden (Milberg et al. 2014). Association between beetle species and oaks was also found by investigating hollow oaks, which are rich in dead branches (Sverdrup-

Thygeson et al. 2010). The study identified 62 Red-listed beetle species associated with oaks, including 23 oak specialists amongst the 62 oak-associated beetles. These oak specialists belonged to various families, such as Ptiliidae, Leiodidae, Scydmaenidae, Staphylinidae, Scarabaeidae, Elateridae, Cantharidae, Anobiidae, Lymexylidae, Melyridae, Nitidulidae, Tenebrionidae, Aderidae and Scraptiidae. A study in Germany revealed the host preference of saproxylic beetle communities on logs of 13 tree species, including birch, European ash, aspen, oak and linden, over a period of two years after harvesting (Müller et al. 2015). The analysis of 381 saproxylic beetle species that emerged from the logs showed that European hornbeam (*Carpinus betulus*) was the most preferred tree species, while European ash, Douglas-fir (*Pseudotsuga menziesii*), European larch (*Larix decidua*) and linden (*Tilia* sp.) were the least preferred.

However, it is not always easy to define whether a species living in deadwood depends on wood fibres, fungal hyphae or other factors (Bakke 1999) and identify beetle-tree associations. Despite that, further studies on the diversity of saproxylic beetles in different tree species and countries are necessary to apply the findings to the conservation and enrichment of unique deadwood habitats and their associated beetles.

More than 3600 species of beetles are recorded in Lithuania up to date (Tamatit et al. 2011, Ferenca et al. 2011, Nagrockaitė et al. 2011, Tamutis 2012, Ferenca et al. 2013, Ivinskis et al. 2013, Monsevičius 2013, Ivinskis et al. 2014, Tamutis and Barševskis 2014, Ferenca and Tamutis 2015, Ivinskis et al. 2015, Tamutis et al. 2015, Ferenca et al. 2016, Paukkunen et al. 2016, Ivinskis et al. 2017, Lekoveckaitė et al. 2017, Pacevičius 2017, Ferenca et al. 2018, Lekoveckaitė et al. 2019, Tamutis and Martinaitis 2019, Monsevičius 2020, Monsevičius 2022). Very little research on beetles in Lithuania has been related to dead wood (Ferenca and Tamutis 2011, Ivinskis et al. 2017, Lekoveckaitė et al. 2017, Pacevičius 2017, Ferenca et al. 2018, Lekoveckaitė et al. 2019, Monsevičius 2020). Our study is the first extended research, designed to investigate beetle communities in deciduous tree species dead wood and especially in its early stage of decay.

## Material and methods

From 2018 to 2021, we collected saproxylic beetle fauna in four protected forest areas of Lithuania (Table 1, Fig. 1). The chosen forests are part of the Natura 2000 network where main forest felling is prohibited or limited to low-intensity selective felling, sanitary felling is also restricted and additional uncut trees must be left in clearings. Additionally, dry trees cannot be felled. Dead wood in the studied forests mainly includes dead branches, standing dead trees (snags) and trees felled by the wind.

A total of 54 deciduous wind-felled trees belonging to six different tree species were chosen for the research (Table 1). Tree species were identified, considering the bark of the tree and the general composition of the stand. As beetle species associated with a specific tree species decrease with increasing decomposition degree, we assumed that primary decay stages should host high beetle diversity. Instead of selecting recently deceased trees in the first stage of decay, we chose weakly-decayed trees in the second stage. The

second stage of wood decay was identified, based on loose bark and knife blade penetration of less than 2 cm (Renvall 1995, Palviainen et al. 2008). The diameters of the tree varied from 21 to 53 cm.

Table 1.

Study sites in four different years with six species of tree trunks. The brackets indicate the number of traps used.

	Biržai forest botanical reserve	Būda botanical - zoological reserve	Dubrava reserve area	Punios Šilas strict nature reserve
2018	<i>F. excelsior</i> (3) <i>A. glutinosa</i> (3)	<i>F. excelsior</i> (3) <i>P. tremula</i> (3)		
2019	<i>F. excelsior</i> (3) <i>A. glutinosa</i> (3)	<i>F. excelsior</i> (3) <i>P. tremula</i> (3)		
2020		<i>Q. robur</i> (3) <i>T. cordata</i> (3)		<i>Q. robur</i> (3) <i>T. cordata</i> (3)
2021		<i>A. glutinosa</i> (3) <i>Betula</i> sp. (3)	<i>P. tremula</i> (3) <i>Betula</i> sp. (3)	<i>Q. robur</i> (3) <i>Betula</i> sp. (3)

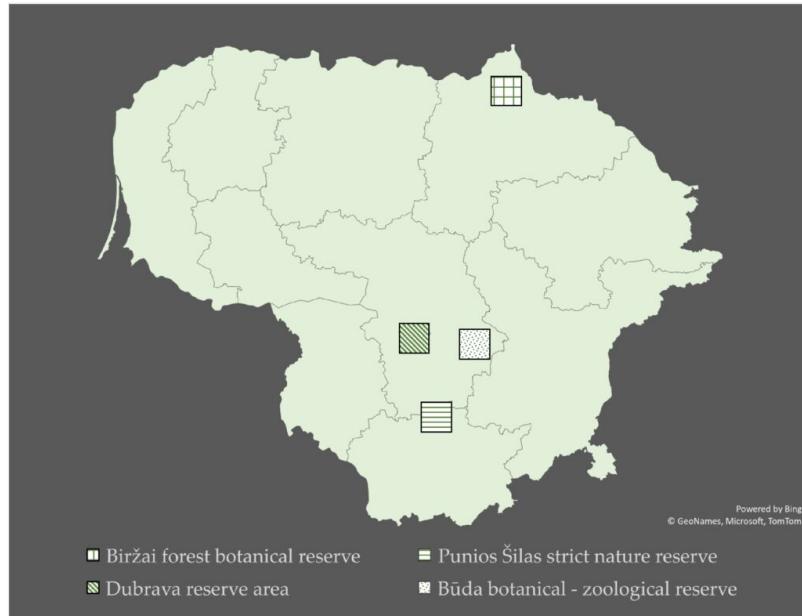


Figure 1. doi

Map of Lithuania with marked research locations.

We used a modified trunk-emergence trap model to collect beetle specimens (Halme et al. 2013) (Fig. 2). Traps were sewn from transparent, air-permeable polyester cloth, to maintain the microclimatic conditions inside the trap unaltered. All traps were designed to cover a 1-metre section of the wind-felled tree and one vertical wall of the trap is longer

than the other to make the insects emerging from the wood fly to the highest point. The bottom of each trap was sealed by joining the cloth with contact tape. For the traps to be properly installed, the trunk must be raised off the ground. We installed traps on the middle parts of such trunks, stretching their walls with the help of sewn ropes so that the material does not form wrinkles. A two-piece collecting jar was attached to the highest point of the trap and filled with > 99% propylene glycol and emptied every two weeks from June to October. In total, we took 82 samples during the four years.



Figure 2. [doi](#)

Modified trunk-emergence trap.

Beetle specimens were identified at the species level. The species names were used following De Jong et al. (2014). The collected material is deposited in the Tadas Ivanauskas Kaunas Zoological Museum and the Vilnius University Life Sciences Center Museum of Zoology.

## Results

A total of 6796 coleopteran specimens belonging to 60 families and 490 species were collected in 54 studied tree trunks of the second stage of wood decay. A small number of

collected specimens were identified to the genus level and regarded as separate species (due to the morphology of specimens) (Table 2).

Table 2.

List of beetle families and species and their abundance collected in emergence traps fixed to horizontal trunks of six different tree species in the secondary stage of decay in Lithuania, June to October, 2018 to 2021: F. ex. – *Fraxinus excelsior*, P. tr. – *Populus tremula*, A. gl. – *Alnus glutinosa*, B. sp. – *Betula* sp., T. co. – *Tilia cordata*, Q. ro. – *Quercus robur*.

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
Aderidae	<sup>1</sup> <i>Phytobaenus amabilis</i> Sahlberg, 1834		1		3		
Anobiidae	<sup>1</sup> <i>Anobium punctatum</i> (De Geer, 1774)			1			
	<sup>1</sup> <i>Dorcatoma (Dorcatoma) dresdensis</i> Herbst, 1792		1				
	<sup>1</sup> <i>Dorcatoma (Pilosodorcatoma) chrysomelina</i> Sturm, 1837				2	1	1
	<sup>1</sup> <i>Ptinus (Bruchoptinus) rufipes</i> Olivier, 1790						3
	<sup>2</sup> <i>Ptinus (Gynopterus) dubius</i> Sturm, 1837						1
	<sup>2</sup> <i>Ptinus (Ptinus) clavipes</i> Panzer, 1806				1	1	
	<sup>1</sup> <i>Ptinus (Ptinus) fur</i> (Linnaeus, 1758)						1
	<sup>2</sup> <i>Ptinus (Ptinus) pilosus</i> Muller, 1821	1					
	<sup>1</sup> <i>Ptinus (Ptinus) subpilosus</i> Sturm, 1837				2	6	12
Anthicidae	<sup>2</sup> <i>Notoxus monoceros</i> (Linnaeus, 1760)					1	
	<sup>2</sup> <i>Omonadus floralis</i> (Linnaeus, 1758)				1		
Anthribidae	<sup>1</sup> <i>Anthribus nebulosus</i> Forster, 1770		1		3	1	
	<sup>1</sup> <i>Dissoleucas niveirostris</i> (Fabricius, 1798)						2
	<sup>1</sup> <i>Platystomos albinus</i> (Linnaeus, 1758)	11	3	1	2	1	1
Apionidae	<sup>3</sup> <i>Betulapion simile</i> (Kirby, 1811)		2		1	1	2
	<sup>3</sup> <i>Catapion pubescens</i> (Kirby, 1811)		2				
	<sup>3</sup> <i>Catapion seniculus</i> (Kirby, 1808)	3	1			2	1
	<sup>3</sup> <i>Kalcapion pallipes</i> (Kirby, 1808)			1			
	<sup>3</sup> <i>Oxystoma craccae</i> (Linnaeus, 1767)		1				
	<sup>3</sup> <i>Taeniapion urticarium</i> (Herbst, 1784)				1		
Biphyllidae	<sup>1</sup> <i>Diplocoelus fagi</i> Guérin-Méneville, 1838		1				
Byrrhidae	<sup>4</sup> <i>Byrrhus (Byrrhus) pilula</i> (Linnaeus, 1758)						1
Cantharidae	<sup>2</sup> <i>Cantharis (Cantharis) nigricans</i> Muller, 1766	1					2
	<sup>1</sup> <i>Malthinus facialis</i> Thomson, 1864				1		
	<sup>1</sup> <i>Malthinus flaveolus</i> (Herbst, 1786)	3			3		
	<sup>1</sup> <i>Malthodes (Malthodes) crassicornis</i> (Mäklin, 1846)					1	1
	<sup>1</sup> <i>Malthodes (Malthodes) fuscus</i> (Waltl, 1838)	2					

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Malthodes (Malthodes) guttifer</i> Kiesenwetter, 1852			4	4		1
	<sup>1</sup> <i>Malthodes (Malthodes) marginatus</i> (Latreille, 1806)	1		1	4	2	2
	<sup>1</sup> <i>Malthodes (Malthodes) minimus</i> (Linnaeus, 1758)	3		1	1		1
	<i>Malthodes</i> sp. Kiesenwetter, 1852	5	3	3			
	<sup>2</sup> <i>Podistra (Absidia) rufotestacea</i> (Letzner, 1845)	1					
	<sup>2</sup> <i>Rhagonycha (Rhagonycha) fulva</i> (Scopoli, 1763)				1		1
	<sup>2</sup> <i>Rhagonycha (Rhagonycha) testacea</i> (Linnaeus, 1758)	2	4	1			1
Carabidae	<sup>2</sup> <i>Agonum (Europilus) thoreyi</i> Dejean, 1828		1				
	<sup>2</sup> <i>Amara (Amara) aenea</i> (De Geer, 1774)		1				
	<sup>2</sup> <i>Amara (Amara) familiaris</i> (Duftschmid, 1812)					3	1
	<sup>2</sup> <i>Calathus (Amphyginus) rotundicollis</i> Dejean, 1828						1
	<sup>2</sup> <i>Carabus (Carabus) granulatus</i> Linnaeus, 1758	3	6	5	9	1	1
	<sup>2</sup> <i>Cychrus caraboides</i> (Linnaeus, 1758)		1				
	<sup>2</sup> <i>Dromius (Dromius) quadraticollis</i> Morawitz, 1862						1
	<sup>1</sup> <i>Dyschiriodes (Eudyschirius) globosus</i> (Herbst, 1783)			1	2		
	<sup>2</sup> <i>Leistus (Leistus) piceus</i> Frölich, 1799	6			1		
	<sup>2</sup> <i>Leistus (Leistus) terminatus</i> (Panzer, 1793)			1	2		1
	<sup>1</sup> <i>Limodromus assimilis</i> (Paykull, 1790)		3	1	1	1	
	<sup>2</sup> <i>Loricera pilicornis</i> (Fabricius, 1775)						1
	<sup>2</sup> <i>Nebria (Nebria) brevicollis</i> (Fabricius, 1792)					2	
	<sup>1</sup> <i>Pterostichus (Bothriopterus) oblongopunctatus</i> (Fabricius, 1787)		1		3		2
	<sup>2</sup> <i>Pterostichus (Eosteropus) aethiops</i> (Panzer, 1796)			1			
	<sup>2</sup> <i>Pterostichus (Morphosoma) melanarius</i> (Illiger, 1798)		5	1			
	<sup>2</sup> <i>Pterostichus (Platysma) niger</i> (Schaller, 1783)	1	2				
	<sup>2</sup> <i>Pterostichus (Pseudomaseus) minor</i> (Gyllenhal, 1827)						1
Cerambycidae	<sup>1</sup> <i>Alosterna tabacicolor</i> (De Geer, 1775)		1			8	2
	<sup>1</sup> <i>Leiopus linnei</i> Wallin, Nylander & Kvamme, 2009						4
	<sup>1</sup> <i>Leiopus nebulosus</i> (Linnaeus, 1758)		1	4			1
	<sup>1</sup> <i>Leptura quadrifasciata</i> Linnaeus, 1758		1				1
	<sup>1</sup> <i>Pogonocherus hispidus</i> (Linnaeus, 1758)					1	
	<sup>1</sup> <i>Pyrrhidium sanguineum</i> (Linnaeus, 1758)						8
	<sup>1</sup> <i>Rhagium (Megarhagium) mordax</i> (De Geer, 1775)	9	16	7	13	11	20
	<sup>1</sup> <i>Rhagium (Rhagium) inquisitor</i> Linnaeus, 1758	1					
	<sup>1</sup> <i>Saperda perforata</i> (Pallas, 1773)		1				
	<sup>1</sup> <i>Saperda scalaris</i> (Linnaeus, 1758)			4	1		3

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Stictoleptura rubra</i> (Linnaeus, 1758)		1				
	<sup>1</sup> <i>Xylotrechus rusticus</i> (Linnaeus, 1758)				1		
Cerylonidae	<sup>1</sup> <i>Cerylon deplanatum</i> Gyllenhal, 1827		3	2	1	2	6
	<sup>1</sup> <i>Cerylon fagi</i> Brisout de Barnevile, 1867	1		2	2	1	1
	<sup>1</sup> <i>Cerylon ferrugineum</i> Stephens, 1830	1			6		1
	<sup>1</sup> <i>Cerylon histeroides</i> (Fabricius, 1792)	1	1	2			2
Chrysomelidae	<sup>3</sup> <i>Batophila rubi</i> (Paykull, 1799)			1			
	<sup>3</sup> <i>Cassida (Cassida) nebulosa</i> Linnaeus, 1758	1					
	<sup>3</sup> <i>Chaetocnema hortensis</i> (Geoffroy, 1785)	1	1				
	<sup>3</sup> <i>Chaetocnema picipes</i> Stephens, 1831				1		
	<sup>3</sup> <i>Cryptocephalus (Burlinius) rufipes</i> (Goeze, 1777)	1					
	<sup>3</sup> <i>Galeruca (Galeruca) tanaceti</i> (Linnaeus, 1758)			1			
	<sup>3</sup> <i>Longitarsus melanocephalus</i> (De Geer, 1775)				1	1	
	<sup>3</sup> <i>Lythraria salicariae</i> (Paykull, 1800)						1
	<sup>3</sup> <i>Oulema melanopus</i> (Linnaeus, 1758)			1	1		
	<sup>3</sup> <i>Phaedon (Phaedon) cochleariae</i> (Fabricius, 1792)	1	1				
	<sup>3</sup> <i>Phyllobrotica quadrimaculata</i> (Linnaeus, 1758)				1		
	<sup>3</sup> <i>Phyllotreta atra</i> (Fabricius, 1775)		2		1		
	<sup>3</sup> <i>Phyllotreta nemorum</i> (Linnaeus, 1758)			1			
	<sup>3</sup> <i>Phyllotreta striolata</i> (Fabricius, 1803)	1	1				
	<sup>3</sup> <i>Phyllotreta undulata</i> Kutschera, 1860	1				1	
	<sup>3</sup> <i>Phyllotreta vittula</i> (Redtenbacher, 1849)	1	7	1		1	1
	<sup>3</sup> <i>Psylliodes (Psylliodes) napi</i> (Fabricius, 1792)						3
Ciidae	<sup>1</sup> <i>Cis alter</i> Silfverberg, 1991				1		
	<sup>1</sup> <i>Cis glabratus</i> Mellié, 1848						1
	<sup>1</sup> <i>Cis jacquemartii</i> Mellié, 1848				4		2
	<sup>1</sup> <i>Cis boleti</i> (Scopoli, 1763)				3		
	<sup>1</sup> <i>Cis castaneus</i> Mellie, 1848						1
	<sup>1</sup> <i>Cis micans</i> (Fabricius, 1792)	1					1
	<sup>1</sup> <i>Ennearthron cornutum</i> (Gyllenhal, 1827)		1				
	<sup>1</sup> <i>Orthocis festivus</i> (Panzer, 1793)				1		
Cleridae	<sup>1</sup> <i>Thanasimus formicarius</i> (Linnaeus, 1758)	7		2	1		4
Coccinellidae	<sup>3</sup> <i>Calvia decemguttata</i> (Linnaeus, 1758)				1		
	<sup>3</sup> <i>Chilocorus renipustulatus</i> (Scriba, 1790)		1				
	<sup>3</sup> <i>Propylea quatuordecimpunctata</i> (Linnaeus, 1758)	1	1			1	1
	<sup>3</sup> <i>Scymnus (Pullus) suturalis</i> Thunberg, 1795			1			

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>3</sup> <i>Tytthaspis sedecimpunctata</i> (Linnaeus, 1758)						1
Corylophidae	<sup>1</sup> <i>Orthoperus punctatus</i> (Wankowicz, 1865)						2
	<sup>1</sup> <i>Orthoperus atomus</i> (Gyllenhal, 1808)	1		1	2	1	8
	<sup>1</sup> <i>Orthoperus rogeri</i> Kraatz, 1874			2			
	<sup>2</sup> <i>Sericoderus lateralis</i> (Gyllenhal, 1827)			1	1	3	
Cryptophagidae	<sup>1</sup> <i>Atomaria procerula</i> Erichson, 1846	1					
	<sup>7</sup> <i>Atomaria pusilla</i> (Paykull, 1798)						1
	<sup>1</sup> <i>Atomaria (Agathengis) nigrirostris</i> Stephens, 1830	2	4	1			2
	<sup>7</sup> <i>Atomaria (Atomaria) fuscipes</i> (Gyllenhal, 1808)	1					
	<sup>1</sup> <i>Atomaria (Atomaria) turgida</i> Erichson, 1846			1			
	<i>Atomaria</i> sp. Stephens, 1829	1	1	2	1		2
	<sup>1</sup> <i>Caenoscelis ferruginea</i> (Sahlberg, 1820)			1			1
	<sup>1</sup> <i>Caenoscelis subdeplanata</i> Brisout de Barneville, 1882			1			
	<sup>1</sup> <i>Cryptophagus dorsalis</i> Sahlberg, 1819			1			
	<sup>1</sup> <i>Cryptophagus badius</i> Sturm, 1845	3			2		
	<sup>1</sup> <i>Cryptophagus dentatus</i> (Herbst, 1793)			1	1	2	
	<sup>1</sup> <i>Cryptophagus fuscicornis</i> Sturm, 1845	1					
	<i>Cryptophagus</i> sp. Herbst, 1792	2	1			1	
	<sup>1</sup> <i>Cryptophagus pallidus</i> Sturm, 1845	4					1
	<sup>1</sup> <i>Cryptophagus pilosus</i> Gyllenhal, 1827	5	1		5		5
	<sup>2</sup> <i>Cryptophagus setulosus</i> Sturm, 1845			1			
	<sup>2</sup> <i>Ephistemus globulus</i> (Paykull, 1798)			4		19	10
	<sup>1</sup> <i>Ephistemus reitteri</i> Casey, 1900			1			
	<sup>1</sup> <i>Micrambe abietis</i> (Paykull, 1798)	4		5			
Cucujidae	<sup>1</sup> <i>Cucujus cinnaberinus</i> (Scopoli, 1763)		13		2	1	5
Curculionidae	<sup>1</sup> <i>Acalles (Acalles) camelus</i> (Fabricius, 1792)				1		
	<sup>3</sup> <i>Brachysomus echinatus</i> (Bonsdorff, 1785)		5		3	2	5
	<sup>3</sup> <i>Ceutorhynchus napi</i> Gyllenhal, 1837		1				
	<sup>3</sup> <i>Ceutorhynchus pallidactylus</i> (Marsham, 1802)						2
	<sup>1</sup> <i>Crypturgus cinereus</i> (Herbst, 1793)		4				
	<sup>1</sup> <i>Crypturgus hispidulus</i> Thomson, 1870		1			3	
	<sup>1</sup> <i>Crypturgus pusillus</i> (Gyllenhal, 1813)				34	2	21
	<sup>3</sup> <i>Curculio (Curculio) glandium</i> Marsham, 1802						1
	<sup>3</sup> <i>Curculio (Curculio) nucum</i> Linnaeus, 1758						1
	<sup>1</sup> <i>Dryocoetes alni</i> (Georg, 1856)		6	1		1	
	<sup>1</sup> <i>Dryocoetes autographus</i> (Ratzeburg, 1837)		217	3		2	1

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Dryocoetes villosus</i> (Fabricius, 1792)						1
	<sup>1</sup> <i>Ernopus tiliae</i> (Panzer, 1793)						1
	<sup>1</sup> <i>Hylastes ater</i> (Paykull, 1800)		1		2	1	
	<sup>1</sup> <i>Hylesinus crenatus</i> (Fabricius, 1787)	94				59	59
	<sup>1</sup> <i>Hylobius (Callirus) abietis</i> (Linnaeus, 1758)				3		
	<sup>3</sup> <i>Hypera (Hypera) postica</i> (Gyllenhal, 1813)				1		
	<sup>3</sup> <i>Otiorhynchus (Chollisanus) raucus</i> (Fabricius, 1777)		7				
	<sup>3</sup> <i>Otiorhynchus (Nihus) scaber</i> (Linnaeus, 1758)	2		1			
	<sup>3</sup> <i>Phyllobius (Dieletus) argentatus</i> (Linnaeus, 1758)				3	12	2
	<sup>3</sup> <i>Phyllobius (Metaphyllum) glaucus</i> (Scopoli, 1763)					2	
	<sup>1</sup> <i>Pityogenes chalcographus</i> (Linnaeus, 1761)	2					
	<sup>3</sup> <i>Polydrusus (Eustolus) corruscus</i> Germar, 1824		1				
	<sup>3</sup> <i>Sciaphilus asperatus</i> (Bonsdorff, 1785)	1			3	27	7
	<sup>1</sup> <i>Scolytus ratzeburgii</i> Janson, 1856				10		
	<sup>3</sup> <i>Stereonychus fraxini</i> (De Geer, 1775)					1	
	<sup>3</sup> <i>Strophosoma (Strophosoma) capitatum</i> (De Geer, 1775)		5		28	14	28
	<sup>1</sup> <i>Taphrorychus bicolor</i> (Herbst, 1793)	2	23			1	
	<sup>1</sup> <i>Trachodes (Trachodes) hispidus</i> (Linnaeus, 1758)		3	1	2	1	1
	<sup>1</sup> <i>Trypodendron domesticum</i> (Linnaeus, 1758)		2	7			
	<sup>1</sup> <i>Trypodendron lineatum</i> (Olivier, 1795)		2	45	15		
	<sup>1</sup> <i>Trypodendron signatum</i> (Fabricius, 1787)	77	16	718	401	6	7
	<sup>1</sup> <i>Xyleborinus saxesenii</i> (Ratzeburg, 1837)					1	2
	<sup>1</sup> <i>Xyleborus cryptographus</i> (Ratzeburg, 1837)		62				
	<sup>1</sup> <i>Xyleborus dispar</i> (Fabricius, 1792)	77	166	54	1	6	9
Dasytidae	<sup>1</sup> <i>Dasytes (Dasytes) niger</i> (Linnaeus, 1761)					1	
	<sup>1</sup> <i>Dasytes (Mesodasytes) plumbeus</i> (Muller, 1776)					1	
Dermestidae	<sup>5</sup> <i>Anthrenus (Florilinus) museorum</i> (Linnaeus, 1761)						1
	<sup>6</sup> <i>Dermestes (Dermestes) ater</i> De Geer, 1774					1	
	<sup>5</sup> <i>Dermestes (Dermestinus) murinus</i> Linnaeus, 1758					1	
Elateridae	<sup>1</sup> <i>Ampedus (Ampedus) erythrogonus</i> (Muller, 1821)				1		1
	<sup>1</sup> <i>Ampedus (Ampedus) nigrinus</i> (Herbst, 1784)	4	3		1		1
	<sup>1</sup> <i>Ampedus (Ampedus) pomorum</i> (Herbst, 1784)	5	5		9		3
	<sup>1</sup> <i>Anostirus castaneus</i> (Linnaeus, 1758)					1	1
	<sup>1</sup> <i>Athous (Athous) haemorrhoidalis</i> (Fabricius, 1801)	1					
	<sup>2</sup> <i>Athous (Athous) vittatus</i> (Gmelin, 1790)						1

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Athous (Haplathous) subfuscus</i> (Muller, 1764)	3		1			
	<sup>1</sup> <i>Dalopius marginatus</i> (Linnaeus, 1758)	6	7	4	2	7	11
	<sup>1</sup> <i>Denticollis linearis</i> (Linnaeus, 1758)	3	1	3	3	3	3
	<sup>1</sup> <i>Denticollis rubens</i> Piller & Mitterpacher, 1783			1			
	<sup>1</sup> <i>Diacanthous undulatus</i> (De Geer, 1774)		2	2		2	6
	<sup>3</sup> <i>Ectinus aterrimus</i> (Linnaeus, 1761)					1	1
	<sup>2</sup> <i>Hemicrepidius niger</i> (Linnaeus, 1758)						1
	<sup>1</sup> <i>Melanotus (Melanotus) castanipes</i> (Paykull, 1800)	1	2		2	1	1
	<sup>1</sup> <i>Melanotus (Melanotus) villosus</i> (Fourcroy, 1785)			1		1	1
	<sup>2</sup> <i>Selatosomus (Pristilophus) cruciatus</i> (Linnaeus, 1758)						2
Endomychidae	<sup>1</sup> <i>Endomychus coccineus</i> (Linnaeus, 1758)	3	10	4	14	3	3
	<sup>1</sup> <i>Leiastes seminiger</i> (Gyllenhal, 1808)						1
	<sup>1</sup> <i>Mycetina cruciata</i> (Schaller, 1783)				2	1	6
Erotylidae	<sup>1</sup> <i>Dacne (Dacne) bipustulata</i> (Thunberg, 1781)						1
	<sup>1</sup> <i>Triplax russica</i> (Linnaeus, 1758)	1					3
Eucnemidae	<sup>1</sup> <i>Eucnemis capucina</i> Ahrens, 1812				1		
	<sup>1</sup> <i>Hylis procerulus</i> (Mannerheim, 1823)		1		2	1	1
	<sup>1</sup> <i>Isohippis melasoides</i> (Laporte de Castelnau, 1835)		1				
	<sup>1</sup> <i>Microrhagus emyi</i> (Rouget, 1856)	1					
	<sup>1</sup> <i>Microrhagus lepidus</i> Rosenhauer, 1847	1			1	1	1
	<sup>1</sup> <i>Microrhagus pygmæus</i> (Fabricius, 1792)					1	
	<sup>1</sup> <i>Xylophilus testaceus</i> (Herbst, 1806)		1				
Geotrupidae	<sup>2</sup> <i>Anoplotrupes stercorosus</i> (Scriba, 1791)	3	19	1	7	2	
Histeridae	<sup>1</sup> <i>Paromalus (Paromalus) parallelepipedus</i> (Herbst, 1792)						1
Hydraenidae	<sup>2</sup> <i>Hydraena (Hydraena) britteni</i> Joy, 1907				1		
Kateretidae	<sup>3</sup> <i>Brachypterus glaber</i> (Newman, 1834)		1				
	<sup>3</sup> <i>Brachypterus urticae</i> (Fabricius, 1792)						2
Laemophloeidae	<sup>1</sup> <i>Cryptolestes ferrugineus</i> (Stephens, 1831)				1		
	<sup>1</sup> <i>Placonotus testaceus</i> (Fabricius, 1787)	1	1				
Lampyridae	<sup>2</sup> <i>Phosphaenus hemipterus</i> (Goeze, 1777)				1		
Latridiidae	<sup>1</sup> <i>Cartodere (Aridius) nodifer</i> (Westwood, 1839)	1	11	3	4	3	2
	<sup>2</sup> <i>Corticaria ferruginea</i> Marsham, 1802		1				2
	<sup>1</sup> <i>Corticaria fulva</i> (Comolli, 1837)		1				
	<sup>1</sup> <i>Corticaria longicollis</i> (Zetterstedt, 1838)				1	1	
	<sup>2</sup> <i>Corticaria longicornis</i> (Herbst, 1783)					2	
	<sup>1</sup> <i>Corticaria serrata</i> (Paykull, 1798)						1

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Corticarina minuta</i> (Fabricius, 1792)	6	8		2		1
	<i>Corticarina</i> sp. Reitter, 1880	1					
	<sup>1</sup> <i>Corticarina similata</i> (Gyllenhal, 1827)			2	14	1	12
	<sup>2</sup> <i>Corticarina truncatella</i> (Mannerheim, 1844)				1		
	<sup>1</sup> <i>Cortinicara gibbosa</i> (Herbst, 1793)	18	7	4	7	25	37
	<sup>1</sup> <i>Enicmus fungicola</i> Thomson, 1868		2		2	1	2
	<sup>1</sup> <i>Enicmus rugosus</i> (Herbst, 1793)	2	1		9	1	12
	<sup>1</sup> <i>Enicmus testaceus</i> (Stephens, 1830)				10		6
	<sup>1</sup> <i>Enicmus transversus</i> (Olivier, 1790)		1		1	1	
	<sup>2</sup> <i>Latridius assimilis</i> (Mannerheim, 1844)				1		
	<sup>1</sup> <i>Latridius consimilis</i> (Mannerheim, 1844)				3	1	
	<sup>1</sup> <i>Latridius hirtus</i> (Gyllenhal, 1827)		7		3	2	2
	<sup>1</sup> <i>Latridius minutus</i> (Linnaeus, 1767)	3	9	12	4	3	10
	<sup>2</sup> <i>Latridius porcatus</i> Herbst, 1793				1		
	<sup>2</sup> <i>Melanophthalma (Melanophthalma) transversalis</i> (Gyllenhal, 1827)		1				
	<sup>1</sup> <i>Stephostethus angusticollis</i> (Gyllenhal, 1827)	4		1	3		
	<i>Stephostethus</i> sp. LeConte, 1878			1			
	<sup>1</sup> <i>Stephostethus pandellei</i> (Brisout, 1863)	3	4		2	2	1
	<sup>1</sup> <i>Stephostethus rugicollis</i> (Olivier, 1790)	1	1	1			
	<sup>2</sup> <i>Thes bergrothi</i> (Reitter, 1880)					1	
Leiodidae	<sup>1</sup> <i>Agathidium atrum</i> (Paykull, 1798)						1
	<sup>8</sup> <i>Agathidium laevigatum</i> Erichson, 1845					4	
	<sup>1</sup> <i>Agathidium (Agathidium) pisanum</i> Brisout, 1872		2		3	1	1
	<sup>1</sup> <i>Agathidium (Agathidium) seminulum</i> (Linnaeus, 1758)			2	2	2	1
	<sup>1</sup> <i>Agathidium (Neoceble) confusum</i> Brisout, 1863	1	1			1	
	<sup>8</sup> <i>Agathidium (Neoceble) convexum</i> Sharp, 1866				1		
	<sup>1</sup> <i>Agathidium (Neoceble) nigripenne</i> (Fabricius, 1792)	1	38	6	1		1
	<sup>8</sup> <i>Agathidium (Neoceble) rotundatum</i> (Gyllenhal, 1827)		1		1		1
	<sup>8</sup> <i>Agathidium (Neoceble) varians</i> Beck, 1817		3				1
	<sup>1</sup> <i>Anisotoma castanea</i> (Herbst, 1792)		1	1		2	
	<sup>1</sup> <i>Anisotoma glabra</i> (Fabricius, 1792)	1	3				2
	<sup>1</sup> <i>Anisotoma humeralis</i> (Fabricius, 1792)	1	1	1	4	2	11
	<sup>1</sup> <i>Anisotoma orbicularis</i> (Herbst, 1792)	1	4	2	2	5	3
	<sup>6</sup> <i>Catops nigrita</i> Erichson, 1837		4	1	4	3	
	<i>Catops</i> sp. Paykull, 1798	1	3				

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>4</sup> <i>Colenis immunda</i> (Sturm, 1807)						1
	<sup>7</sup> <i>Colon (Myloechus) brunneum</i> (Latreille, 1807)						1
	<sup>7</sup> <i>Colon viennense</i> Herbst, 1797					1	
	<sup>8</sup> <i>Fissocatops westi</i> (Krogerus, 1931)			3	1		
	<sup>8</sup> <i>Hydnobius spinipes</i> (Gyllenhal, 1813)		1				
	<sup>8</sup> <i>Leiodes pallens</i> (Sturm, 1807)					1	
	<i>Leiodes</i> sp. Latreille, 1796					1	
	<sup>1</sup> <i>Liodopria serricornis</i> (Gyllenhal, 1813)				1		
	<sup>8</sup> <i>Nargus (Nargus) velox</i> (Spence, 1815)	1					
	<i>Nargus</i> sp. Thomson, 1867			1			
	<sup>8</sup> <i>Ptomaphagus (Ptomaphagus) varicornis</i> (Rosenhauer, 1847)			1		1	
	<sup>8</sup> <i>Sciadrepoides watsoni</i> (Spence, 1815)	1	2	6		6	
Lucanidae	<sup>1</sup> <i>Dorcus parallelipipedus</i> (Linnaeus, 1785)						1
	<sup>1</sup> <i>Sinodendron cylindricum</i> (Linnaeus, 1758)					1	
Lycidae	<sup>1</sup> <i>Lygistopterus sanguineus</i> (Linnaeus, 1758)						3
Lymexylidae	<sup>1</sup> <i>Hylecoetus dermestoides</i> (Linnaeus, 1861)			38	52	5	2
	<sup>1</sup> <i>Lymexylon navale</i> (Linnaeus, 1758)					1	
Malachiidae	<sup>1</sup> <i>Malachius bipustulatus</i> (Linnaeus, 1758)					1	2
Melandryidae	<sup>1</sup> <i>Hypulus quercinus</i> (Quensel, 1790)						5
	<sup>1</sup> <i>Melandrya dubia</i> (Schaller, 1783)	1	2		2	1	3
	<sup>1</sup> <i>Orchesia (Clinocara) undulata</i> Kraatz, 1853	7	8	8	4	2	8
	<sup>1</sup> <i>Orchesia (Orchesia) micans</i> (Panzer, 1794)		1		1		2
Melolonthidae	<sup>1</sup> <i>Serica brunnea</i> (Linnaeus, 1758)		1	3	3	3	4
Monotomidae	<sup>3</sup> <i>Monotoma (Monotoma) picipes</i> Herbst, 1793		1				
	<sup>1</sup> <i>Rhizophagus fenestralis</i> (Linnaeus, 1758)	10	20	3	8	13	5
	<sup>1</sup> <i>Rhizophagus (Anomophagus) puncticollis</i> Sahlberg, 1837	1					
	<sup>1</sup> <i>Rhizophagus (Cyanostolus) aeneus</i> Richter, 1820						2
	<sup>1</sup> <i>Rhizophagus (Rhizophagus) bipustulatus</i> (Fabricius, 1792)	9	3	5	4	4	4
	<sup>1</sup> <i>Rhizophagus (Rhizophagus) dispar</i> (Paykull, 1800)	11	3	30		5	1
	<sup>1</sup> <i>Rhizophagus (Rhizophagus) ferrugineus</i> (Paykull, 1800)			1			
	<sup>1</sup> <i>Rhizophagus (Rhizophagus) nitidulus</i> (Fabricius, 1798)	1	1	3			
	<sup>1</sup> <i>Rhizophagus (Rhizophagus) oblongicollis</i> Blatch & Horner, 1892			2			

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
Mordellidae	<sup>1</sup> <i>Mordellistena (Mordellistena) humeralis</i> (Linnaeus, 1758)				1		
	<sup>1</sup> <i>Tomoxia bucephala</i> (Costa, 1854)		2		1		1
Mycetophagidae	<sup>1</sup> <i>Litargus (Litargus) connexus</i> (Geoffroy, 1785)				1		3
	<sup>1</sup> <i>Mycetophagus (Mycetophagus) quadripustulatus</i> (Linnaeus, 1761)				1		
	<sup>1</sup> <i>Mycetophagus (Mycetoxides) fulvicollis</i> Fabricius, 1793						1
	<sup>1</sup> <i>Mycetophagus (Philomyces) populi</i> Fabricius, 1798				1		
	<sup>1</sup> <i>Mycetophagus (Ulolendus) atomarius</i> (Fabricius, 1787)						3
	<sup>1</sup> <i>Mycetophagus (Ulolendus) piceus</i> (Fabricius, 1777)						1
Nemonychidae	<sup>3</sup> <i>Cimberis attelaboides</i> (Fabricius, 1787)				1		
Nitidulidae	<sup>3</sup> <i>Brassicogethes aeneus</i> (Fabricius, 1775)	1	1			1	1
	<sup>3</sup> <i>Brassicogethes subaeneus</i> (Sturm, 1845)	1	1				
	<sup>2</sup> <i>Carpophilus ligneus</i> Murray, 1864						1
	<sup>1</sup> <i>Cychramus luteus</i> (Fabricius, 1787)	3	2	10	3	38	5
	<sup>1</sup> <i>Cychramus variegatus</i> (Herbst, 1792)	4	3	3	4	7	3
	<sup>1</sup> <i>Cylloides ater</i> (Herbst, 1792)						1
	<sup>1</sup> <i>Epuraea angustula</i> Sturm, 1844	1		1			
	<sup>1</sup> <i>Epuraea marseuli</i> Reitter, 1872	10		11	1		
	<sup>2</sup> <i>Epuraea melanocephala</i> (Marsham, 1802)						1
	<sup>1</sup> <i>Epuraea neglecta</i> (Heer, 1841)	6	21	10	5		4
	<sup>1</sup> <i>Epuraea oblonga</i> (Herbst, 1793)			1			
	<sup>1</sup> <i>Epuraea pallescens</i> (Stephens, 1835)	3	3	3	2		
	<sup>1</sup> <i>Epuraea unicolor</i> (Olivier, 1790)	10		2			
	<sup>1</sup> <i>Epuraea variegata</i> (Herbst, 1793)	8	7	3	8		1
	<sup>1</sup> <i>Glischrochilus hortensis</i> (Geoffroy in Fourcroy, 1785)	93	51	11	6	1	6
	<sup>1</sup> <i>Glischrochilus quadriguttatus</i> (Fabricius, 1776)	2	21		2		14
	<sup>1</sup> <i>Glischrochilus quadripunctatus</i> (Linnaeus, 1758)		1				
	<sup>1</sup> <i>Glischrochilus quadrisignatus</i> (Say, 1835)				2		
	<sup>1</sup> <i>Ipidia binotata</i> Reitter, 1875			1	2		2
	<sup>3</sup> <i>Lamiogethes brunnicornis</i> (Sturm, 1845)						1
	<sup>1</sup> <i>Soronia punctatissima</i> (Illiiger, 1794)	1		1			
Phalacridae	<sup>3</sup> <i>Olibrus affinis</i> (Sturm, 1807)						3
	<sup>3</sup> <i>Olibrus aeneus</i> (Fabricius, 1792)						1
	<sup>3</sup> <i>Stilbus testaceus</i> (Panzer, 1797)						1
	<sup>3</sup> <i>Stilbus oblongus</i> (Erichson, 1845)		4				

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
Ptiliidae	<sup>2</sup> <i>Acrotrichis (Acrotrichis) sitkaensis</i> (Motschulsky, 1845)			3			
	<i>Acrotrichis</i> sp. Motschulsky, 1848	1	11	8			
	<sup>1</sup> <i>Baeocera variolosa</i> (Mulsant & Rey, 1873)	9	4	1			
	<sup>2</sup> <i>Nephantes titan</i> (Newman, 1834)	1					
	<sup>1</sup> <i>Ptenidium (Gillmeisterium) nitidum</i> (Heer, 1841)			1		1	
	<sup>1</sup> <i>Ptenidium (Matthewsium) turgidum</i> Thomson, 1855		1				
	<sup>1</sup> <i>Ptenidium (Ptenidium) pusillum</i> (Gyllenhal, 1808)			1		2	
	<sup>1</sup> <i>Ptinella aptera</i> (Guérin-Méneville, 1839)	14	1				
	<sup>1</sup> <i>Ptinella limbata</i> (Heer, 1841)	5	3		1	1	
Pyrochroidae	<sup>1</sup> <i>Pyrochroa coccinea</i> (Linnaeus, 1761)	3	1		5		9
	<sup>1</sup> <i>Schizotus pectinicornis</i> (Linnaeus, 1758)	5	1		2	8	10
Salpingidae	<sup>1</sup> <i>Salpingus planirostris</i> (Fabricius, 1787)				1	1	1
	<sup>1</sup> <i>Salpingus ruficollis</i> (Linnaeus, 1761)	13	2	11	2	2	2
	<sup>1</sup> <i>Vincenzellus ruficollis</i> (Panzer, 1794)					1	
Scirtidae	<sup>2</sup> <i>Cyphon ochraceus</i> Stephens, 1830	18	16	306	66	2	
	<sup>2</sup> <i>Cyphon padi</i> (Linnaeus, 1758)		2		2		1
	<sup>2</sup> <i>Microcara testacea</i> (Linnaeus, 1767)	1		13	4		
Scaptiidae	<sup>1</sup> <i>Anaspis brunnipes</i> (Mulsant, 1856)						1
	<sup>1</sup> <i>Anaspis (Anaspis) frontalis</i> (Linnaeus, 1758)		3	1	2	1	2
	<sup>1</sup> <i>Anaspis (Anaspis) thoracica</i> (Linnaeus, 1758)	7	2	2	2	2	17
	<sup>1</sup> <i>Scaptia fuscula</i> Muller, 1821						1
Scydmaenidae	<sup>1</sup> <i>Microscydmus (Microscydmus) nanus</i> (Schaum, 1844)						1
	<sup>1</sup> <i>Neuraphes elongatus</i> (Müller & Kunze, 1822)				1		
	<sup>1</sup> <i>Scydmaenus (Parallomicrus) rufus</i> Muller & Kunze, 1822	1					
	<sup>1</sup> <i>Scydmoraphes minutus</i> (Chaudoir, 1845)						1
	<sup>1</sup> <i>Stenichnus (Cyrtoscydmus) collaris</i> (Muller & Kunze, 1822)				1	2	
	<sup>1</sup> <i>Stenichnus (Cyrtoscydmus) godarti</i> (Latreille, 1806)				1	3	6
	<sup>1</sup> <i>Stenichnus (Cyrtoscydmus) scutellaris</i> (Muller & Kunze, 1822)			1	2		1
Silphidae	<sup>6</sup> <i>Nicrophorus vespillo</i> (Linnaeus, 1758)				1		
	<sup>6</sup> <i>Nicrophorus vespilloides</i> Herbst, 1783	1	5		2		
	<sup>6</sup> <i>Oiceoptoma thoracicum</i> (Linnaeus, 1758)	1					
	<sup>6</sup> <i>Phosphuga atrata</i> (Linnaeus, 1758)	1	4	1	1	2	2
	<sup>6</sup> <i>Silpha carinata</i> Herbst, 1783					2	2
Silvanidae	<sup>1</sup> <i>Dendrophagus crenatus</i> (Paykull, 1799)	2			1		

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Silvanoprus fagi</i> (Guérin-Méneville, 1844)		1				1
	<sup>1</sup> <i>Silvanus bidentatus</i> (Fabricius, 1792)		1				
	<sup>1</sup> <i>Silvanus unidentatus</i> (Olivier, 1790)	1					
Sphindidae	<sup>1</sup> <i>Aspidiphorus orbiculatus</i> (Gyllenhal, 1808)	5	98	4	50	35	12
Staphylinidae	<sup>2</sup> <i>Acrotona sylvicola</i> (Kraatz, 1856)		5				
	<sup>1</sup> <i>Acrulia inflata</i> (Gyllenhal, 1813)	1		7			
	<sup>2</sup> <i>Aleochara erythroptera</i> Gravenhorst, 1806			4			
	<sup>7</sup> <i>Aleochara fumata</i> Gravenhorst, 1802					1	
	<sup>1</sup> <i>Aleochara sparsa</i> Heer, 1839		14				
	<sup>2</sup> <i>Aleochara (Euryodma) brevipennis</i> Gravenhorst, 1806	1	2				
	<sup>2</sup> <i>Alevonota gracilenta</i> (Erichson, 1839)						1
	<sup>2</sup> <i>Aloconota gregaria</i> (Erichson, 1839)						4
	<sup>5</sup> <i>Amidobia talpa</i> (Heer, 1841)					3	
	<sup>2</sup> <i>Amischa analis</i> (Gravenhorst, 1802)		4				
	<sup>1</sup> <i>Anomognathus cuspidatus</i> (Erichson, 1839)					1	
	<sup>2</sup> <i>Anotylus rugosus</i> (Fabricius, 1775)		2				
	<sup>2</sup> <i>Atheta amicula</i> (Stephens, 1832)					3	
	<sup>1</sup> <i>Atheta atramentaria</i> (Gyllenhal, 1810)	3					
	<sup>8</sup> <i>Atheta boleticola</i> J. Sahlberg, 1876	10	5	12			
	<sup>1</sup> <i>Atheta crassicornis</i> (Fabricius, 1792)	3	4	55	23	5	
	<sup>5</sup> <i>Atheta divisa</i> (Märkel, 1844)	8	1				2
	<sup>7</sup> <i>Atheta fungi</i> (Gravenhorst, 1806)	14	6	9	10	1	5
	<sup>1</sup> <i>Atheta laticollis</i> (Stephens, 1832)						2
	<sup>2</sup> <i>Atheta longicornis</i> (Gravenhorst, 1802)					3	
	<sup>1</sup> <i>Atheta nigritula</i> (Gravenhorst, 1802)	8	7	19		1	
	<i>Atheta</i> sp. Thomson, 1858	41	7	2		4	5
	<sup>1</sup> <i>Atheta (Atheta) hypnorum</i> (Kiesenwetter, 1850)		1			8	3
	<sup>1</sup> <i>Atrecus affinis</i> (Paykull, 1789)	2	5				
	<sup>1</sup> <i>Batrisodes delaporti</i> (Aube, 1833)			1			
	<sup>1</sup> <i>Batrisodes hubenthali</i> Reitter, 1913						4
	<i>Batrisodes</i> sp. Reitter, 1882		1				
	<sup>1</sup> <i>Batrisodes venustus</i> (Reichenbach, 1816)						1
	<sup>1</sup> <i>Bibloporus bicolor</i> (Denny, 1825)	3	4		2		1
	<sup>2</sup> <i>Bisnius firmarius</i> (Gravenhorst, 1802)	3	1		2		1
	<sup>2</sup> <i>Bisnius nitidulus</i> (Gravenhorst, 1802)	1					
	<sup>2</sup> <i>Bolitobius castaneus</i> (Stephens, 1832)				1		

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Bolitochara lucida</i> (Gravenhorst, 1802)				1		2
	<sup>1</sup> <i>Bolitochara mulsanti</i> Sharp, 1875		2				
	<sup>1</sup> <i>Bolitochara obliqua</i> Erichson, 1837	25	40	15	26	4	32
	<sup>1</sup> <i>Bolitochara pulchra</i> (Gravenhorst, 1806)		4				
	<sup>2</sup> <i>Brachygluta haematica</i> (Reichenbach, 1816)				1		
	<sup>1</sup> <i>Carphecis striatus</i> (Olivier, 1795)		1				
	<sup>1</sup> <i>Dadobia immersa</i> (Erichson, 1837)	2	1		1		
	<sup>1</sup> <i>Dinaraea aequata</i> (Erichson, 1837)		7	5	1		3
	<sup>1</sup> <i>Dinaraea angustula</i> (Gyllenhal, 1810)	4	1		2	4	
	<sup>1</sup> <i>Dinaraea linearis</i> (Gravenhorst, 1802)	4	10				4
	<i>Dinaraea</i> sp. Thomson, 1858			1			
	<i>Euplectus</i> sp. Leach, 1817	1	1				
	<sup>1</sup> <i>Euplectus piceus</i> Motschulsky, 1835			1			5
	<sup>1</sup> <i>Euryusa castanoptera</i> Kraatz, 1856	2	3	4	9	2	2
	<sup>2</sup> <i>Eusphalerum</i> ( <i>Eusphalerum</i> ) <i>minutum</i> (Fabricius, 1792)			1	2		
	<sup>2</sup> <i>Gabrius breviventer</i> (Sperk, 1835)	10		5		1	
	<sup>2</sup> <i>Gabrius nigritulus</i> (Gravenhorst, 1802)		1				
	<sup>2</sup> <i>Gabrius osseticus</i> (Kolenati, 1846)		4				
	<sup>1</sup> <i>Gabrius splendidulus</i> (Gravenhorst, 1802)	3	9	2	1		1
	<sup>1</sup> <i>Geostiba</i> ( <i>Geostiba</i> ) <i>circellaris</i> (Gravenhorst, 1806)		3			5	2
	<sup>7</sup> <i>Gyrophaena gentilis</i> Erichson, 1839				1		
	<sup>7</sup> <i>Gyrophaena joyoides</i> Wüsthoff, 1937					2	
	<sup>1</sup> <i>Haploglossa gentilis</i> (Märkel, 1844)					1	2
	<sup>5</sup> <i>Haploglossa pulla</i> (Gyllenhal, 1827)					1	
	<sup>1</sup> <i>Homalota plana</i> (Gyllenhal, 1810)				2	1	
	<sup>8</sup> <i>Hypnogyra angularis</i> (Ganglbauer, 1895)	1					
	<sup>2</sup> <i>Ilyobates nigricollis</i> (Paykull, 1800)	16					
	<sup>2</sup> <i>Ischnosoma splendidum</i> (Gravenhorst, 1806)			2	1		
	<sup>1</sup> <i>Leptusa pulchella</i> (Mannerheim, 1831)	13		7			
	<sup>4</sup> <i>Lesteva</i> ( <i>Lesteva</i> ) <i>longoelytrata</i> (Goeze, 1777)	1	2				
	<sup>2</sup> <i>Liogluta granigera</i> (Kiesenwetter, 1850)			2			
	<sup>1</sup> <i>Lordithon pulchellus</i> (Mannerheim, 1830)					7	
	<sup>1</sup> <i>Lordithon trinotatus</i> (Erichson, 1839)					3	
	<sup>1</sup> <i>Lordithon lunulatus</i> (Linnaeus, 1760)	1	2		1		1
	<sup>1</sup> <i>Lordithon trimaculatus</i> (Fabricius, 1793)		1				
	<sup>1</sup> <i>Megarthrus depressus</i> (Paykull, 1789)			1			

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Mycetoporus punctus</i> (Gravenhorst, 1806)		1				
	<sup>2</sup> <i>Nehemitropia lividipennis</i> (Mannerheim, 1831)			2	1		
	<sup>1</sup> <i>Nudobius lentsus</i> (Gravenhorst, 1806)		4		2	1	1
	<sup>2</sup> <i>Olophrum assimile</i> (Paykull, 1800)			1			
	<sup>2</sup> <i>Olophrum fuscum</i> (Gravenhorst, 1806)	1					
	<sup>1</sup> <i>Omalium rivulare</i> (Paykull, 1789)	2					
	<sup>1</sup> <i>Othius lapidicola</i> Markel & Kiesenwetter, 1848	1					
	<sup>2</sup> <i>Othius subuliformis</i> Stephens, 1833		4			1	
	<sup>3</sup> <i>Oxypoda acuminata</i> (Stephens, 1832)	6	3			2	3
	<sup>2</sup> <i>Oxypoda annularis</i> (Mannerheim, 1830)				7		
	<sup>3</sup> <i>Oxypoda brevicornis</i> (Stephens, 1832)			24	1		5
	<sup>2</sup> <i>Oxypoda opaca</i> (Gravenhorst, 1802)		2		3	1	3
	<sup>2</sup> <i>Oxypoda praecox</i> Erichson, 1839				3	1	
	<sup>1</sup> <i>Oxypoda (Mycetodrepa) alternans</i> (Gravenhorst, 1802)	3	11	1	1		4
	<i>Oxypoda</i> sp. Mannerheim, 1831			4	1	2	2
	<sup>2</sup> <i>Philhygra elongatula</i> (Gravenhorst, 1802)	4	3		3	3	6
	<sup>2</sup> <i>Philhygra luridipennis</i> (Mannerheim, 1830)				1	1	
	<i>Philhygra</i> sp. Mulsant & Rey, 1873		5				1
	<sup>2</sup> <i>Philonthus decorus</i> (Gravenhorst, 1802)					1	
	<sup>1</sup> <i>Phloeonomus (Phloeonomodes) minimus</i> (Erichson, 1839)						1
	<sup>1</sup> <i>Phloeonomus (Phloeonomus) punctipennis</i> Thomson, 1867	1		2		1	9
	<sup>1</sup> <i>Phloeonomus (Phloeonomus) pusillus</i> (Gravenhorst, 1806)						3
	<sup>1</sup> <i>Phloeopora testacea</i> (Mannerheim, 1830)	1			3		
	<sup>1</sup> <i>Phloeostiba plana</i> (Paykull, 1792)				5		8
	<sup>1</sup> <i>Phyllodrepa (Dropephylla) ioptera</i> (Stephens, 1832)						1
	<sup>1</sup> <i>Phyllodrepa (Phyllodrepa) melanocephala</i> (Fabricius, 1787)		1				
	<sup>1</sup> <i>Phyllodrepaidea crenata</i> (Gravenhorst, 1802)		1				
	<sup>1</sup> <i>Placusa incompleta</i> Sjöberg, 1934					1	
	<sup>1</sup> <i>Placusa (Placusa) atrata</i> (Mannerheim, 1831)	10	1	3			
	<sup>1</sup> <i>Placusa (Placusa) tachyporoides</i> (Waltl, 1838)	21	1				
	<sup>1</sup> <i>Plectophloeus fischeri</i> (Aube, 1833)		1	1	1	2	1
	<sup>1</sup> <i>Plectophloeus nubigena</i> Reitter, 1877						1
	<sup>1</sup> <i>Proteinus brachypterus</i> (Fabricius, 1792)	4					

Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>1</sup> <i>Quedius (Microsaurus) brevicornis</i> (Thomson, 1860)		1				
	<sup>1</sup> <i>Quedius (Microsaurus) cruentus</i> (Olivier, 1795)		6	1	1		
	<sup>5</sup> <i>Quedius (Microsaurus) longicornis</i> Kraatz, 1857	2	2				
	<sup>1</sup> <i>Quedius (Microsaurus) scitus</i> (Gravenhorst, 1806)			1	1		
	<sup>1</sup> <i>Quedius (Microsaurus) xanthopus</i> Erichson, 1839	2	5		3		
	<sup>1</sup> <i>Quedius (Quedionuchus) plagiatus</i> Mannerheim, 1843		1				
	<sup>4</sup> <i>Quedius (Quedius) fuliginosus</i> (Gravenhorst, 1802)					6	
	<sup>4</sup> <i>Quedius (Quedius) molochinus</i> (Gravenhorst, 1806)	2					
	<sup>1</sup> <i>Saulcyella schmidti</i> (Maerkel, 1844)				1		
	<sup>1</sup> <i>Scaphidium quadrimaculatum</i> Olivier, 1790				1		
	<sup>1</sup> <i>Scaphisoma agaricinum</i> (Linnaeus, 1758)	1	14		33	3	15
	<sup>1</sup> <i>Scaphisoma boleti</i> (Panzer, 1793)		1	3		2	
	<sup>1</sup> <i>Sepedophilus bipunctatus</i> (Gravenhorst, 1802)				1		
	<sup>1</sup> <i>Sepedophilus bipustulatus</i> (Gravenhorst, 1802)				15	1	22
	<sup>1</sup> <i>Sepedophilus littoreus</i> (Linnaeus, 1758)	3	10	4	2		1
	<sup>1</sup> <i>Sepedophilus testaceus</i> (Fabricius, 1793)				2		
	<sup>2</sup> <i>Stenus humilis</i> Erichson, 1839				1		
	<sup>2</sup> <i>Stenus bimaculatus</i> Gyllenhal, 1810				1		
	<sup>2</sup> <i>Stenus juno</i> (Paykull, 1789)	1	1				
	<sup>2</sup> <i>Stenus lustrator</i> Erichson, 1839	1					
	<sup>2</sup> <i>Syntomium aeneum</i> (Muller, 1821)		2		1		2
	<sup>2</sup> <i>Tachinus fimetarius</i> Gravenhorst, 1802	2		1			
	<sup>2</sup> <i>Tachinus laticollis</i> Gravenhorst, 1802			1			
	<sup>2</sup> <i>Tachinus marginellus</i> (Fabricius, 1781)		3		1		
	<sup>2</sup> <i>Tachinus signatus</i> Gravenhorst, 1802		1				
	<sup>2</sup> <i>Tachyporus hypnorum</i> (Fabricius, 1775)				1		2
	<sup>4</sup> <i>Tachyporus obtusus</i> (Linnaeus, 1767)					2	1
	<sup>2</sup> <i>Tachyporus pusillus</i> Gravenhorst, 1806						2
	<sup>1</sup> <i>Trimium brevicorne</i> (Reichenbach, 1816)					1	
	<sup>1</sup> <i>Tyrus mucronatus</i> (Panzer, 1805)						1
	<sup>2</sup> <i>Xantholinus linearis</i> (Olivier, 1795)				1		
	<sup>2</sup> <i>Xantholinus (Xantholinus) longiventris</i> Heer, 1839				2		
	<sup>2</sup> <i>Xylodromus depressus</i> (Gravenhorst, 1802)					2	
	<sup>5</sup> <i>Zyras humeralis</i> (Gravenhorst, 1802)	3					
Tenebrionidae	<sup>1</sup> <i>Bolitophagus reticulatus</i> (Linnaeus, 1767)				19		
	<sup>1</sup> <i>Hypophloeus unicolor</i> (Piller & Mitterpacher, 1783)				11	3	

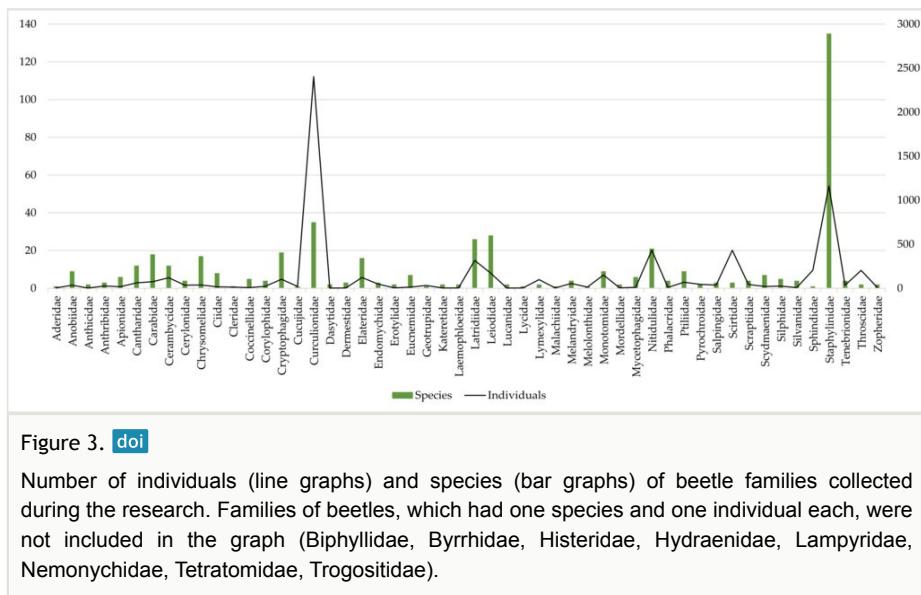
Family	Species	F. ex	P. tr	A. gl	B. sp.	T. co	Q. ro
	<sup>2</sup> <i>Lagria hirta</i> (Linnaeus, 1758)		1				
	<sup>1</sup> <i>Uloma culinaris</i> (Linnaeus, 1758)				1		
Tetratomidae	<sup>1</sup> <i>Hallomenus binotatus</i> (Quensel, 1790)					1	
Throscidae	<sup>2</sup> <i>Trixagus carinifrons</i> (Bonvouloir, 1859)			1		2	1
	<sup>2</sup> <i>Trixagus dermestoides</i> (Linnaeus, 1766)	9	15	11	92	12	60
Trogositidae	<sup>1</sup> <i>Nemozoma elongatum</i> (Linnaeus, 1761)			1			
Zopheridae	<sup>1</sup> <i>Colydium elongatum</i> (Fabricius, 1787)						2
	<sup>1</sup> <i>Synchita humeralis</i> (Fabricius, 1792)	2	1		3		1
<b>Total</b>							
<b>Species</b>		162	205	143	198	159	210
<b>Individuals</b>		956	1383	1686	1347	556	859

<sup>1</sup> Saproxylic species; <sup>2</sup> species related with accumulated organic matter, plant litter, mud and soil; <sup>3</sup> with various plants; <sup>4</sup> with mosses; <sup>5</sup> with nests of bird and other animals; <sup>6</sup> with carrion; <sup>7</sup> with fungal fruiting bodies; <sup>8</sup> habitat association unknown, but considered non-saproxylic.

The most diverse beetle families were Staphylinidae (135 species), Curculionidae (35 species), Leiodidae (27 species) and Latridiidae (26 species) (Fig. 3, Table 2). Together, they represent 45.51% of collected species in dead wood trunks. Sixteen beetle families were represented by a single species each (Aderidae, Biphyllidae, Byrrhidae, Cleridae, Cucujidae, Geotrupidae, Histeridae, Hydraenidae, Lampyridae, Lycidae, Malachiidae, Melolonthidae, Nemonychidae, Sphindidae, Tetratomidae, Trogositidae) (Fig. 3, Table 2). Other beetle families contained from 2 to 21 species (Fig. 3, Table 2).

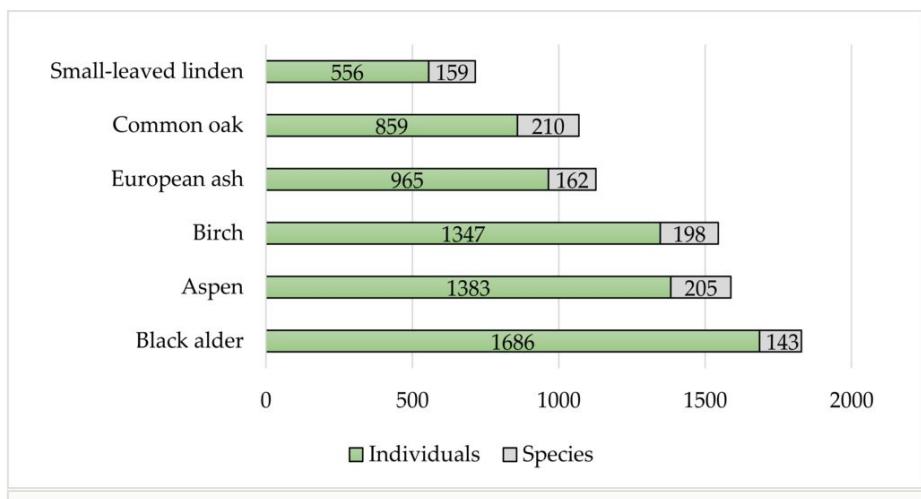
Curculionidae (2403 specimens) and Staphylinidae (1162 specimens) were the most abundant families (Fig. 3, Table 2). Together they represent 52.46% of collected specimens. Eight beetle families (Biphyllidae, Byrrhidae, Histeridae, Hydraenidae, Lampyridae, Nemonychidae, Tetratomidae and Trogositidae) (Fig. 3, Table 2) were represented only by a single specimen, while the majority of beetle families contained from 2 to 433 specimens.

Amongst the species, the most abundant were *Trypodendron signatum* (Fabricius, 1787) (1225 specimens), *Cyphon ochraceus* Stephens, 1830 (408 specimens), *Xyleborus dispar* (Fabricius, 1792) (313 specimens), *Dryocoetes autographus* (Ratzeburg, 1837) (223 specimens), *Hylesinus crenatus* (Fabricius, 1787) (212 specimens), *Aspidiphorus orbiculatus* (Gyllenhal, 1808) (204 specimens), *Trixagus dermestoides* (Linnaeus, 1766) (199 specimens), *Glischrochilus hortensis* (Geoffroy in Fourcroy, 1785) (168 specimens) and *Bolitochara obliqua* Erichson, 1837 (142 specimens). Other beetle species abundance varied from 1 to 98 specimens (Table 2).

Figure 3. [doi](#)

Number of individuals (line graphs) and species (bar graphs) of beetle families collected during the research. Families of beetles, which had one species and one individual each, were not included in the graph (Biphylliidae, Byrrhidae, Histeridae, Hydraenidae, Lampyridae, Nemonychidae, Tetratomidae, Trogositidae).

In total, 965 specimens belonging to 162 beetle species emerged from trunks of European ash, 1383 specimens and 205 species – from trunks of aspen, 1686 specimens and 143 species – from trunks of Black alder, 1347 specimens and 198 species – from trunks of birch, 556 specimens and 159 species – from trunks of small-leaved linden and 859 specimens and 210 species – from trunks of common oak (Fig. 4, Table 2).

Figure 4. [doi](#)

Richness and abundance of beetles in six different tree species. Note: the number of decaying tree trunks sampled with emergence traps was not equal amongst all six tree species. There were 12 trunks sampled from European ash, six from small-leaved linden and nine from each of the remaining species.

Out of the 490 beetle species collected in the research, almost half (246) were found in only one tree species. A total of 85 species were found in two tree species, 58 in three tree species, 46 in four tree species, 28 in five tree species and 27 in all six tree species (Fig. 5). Excluding beetle species with five or fewer specimens, there were 10 beetle species in one tree species, 17 in two tree species, 36 in three tree species, 41 in four tree species, 28 in five tree species and 27 in six tree species (Fig. 5).

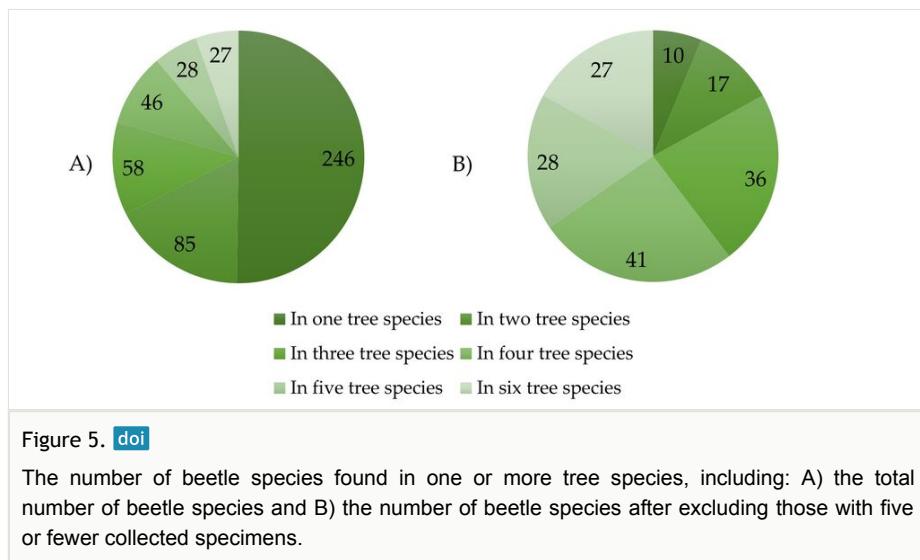


Figure 5. [doi](#)

The number of beetle species found in one or more tree species, including: A) the total number of beetle species and B) the number of beetle species after excluding those with five or fewer collected specimens.

## Discussion

Our research is the first thorough study of saproxylic beetles using the emergence type of traps in Lithuania. It has revealed a huge diversity of saproxylic beetles, accounting for about 13% of all known beetle species in Lithuania. A range of methods is used to collect saproxylic beetles (Ranius and Jansson 2002, Jonsell and Hansson 2007, Peuhu et al. 2019), with window traps, trunk window traps and emergence or elector traps amongst the most popular (Peuhu et al. 2019). A similar method of closed emergence traps for collecting beetles has been used in Germany (Irmler et al. 1996, Müller et al. 2015), France (Bouget et al. 2012), Italy (Parisi et al. 2021) and Sweden (Wikars et al. 2005, Gibb et al. 2006, Hjältén et al. 2010). For example, in Germany, with half the survey time, but with considerably more trunks and almost twice as many tree species, a lower beetle diversity (381 species) was collected compared to our study (Müller et al. 2015).

Of the 490 beetle species we identified, 440 were previously discovered in dead wood of various tree species: European ash, small-leaved linden, common oak, aspen, birch, black alder, Norway spruce (*Picea abies*), pine (*Pinus sylvestris*), common beech (*Fagus sylvatica*), silver fir (*Abies alba*), rowan (*Sorbus aucuparia*) and goat willow (*Salix caprea*), which are the most common tree species in Europe (Kaila 1993, Irmler et al. 1996, Martikainen 2001, Jonsell et al. 2004, Lindhe and Lindelöw 2004, Wikars et al. 2005, Byk

et al. 2006, Gibb et al. 2006, Müller et al. 2007, Djupström et al. 2008, Müller and Bussler 2008, Unal et al. 2009, Hjältén et al. 2010, Horák 2011, Ranius et al. 2011, Bouget et al. 2012, Jonsell 2012, Lassauce et al. 2013, Sawoniewicz 2013, Vindstad et al. 2014, Redolfi De Zan et al. 2014, Papis and Mokrzycki 2015, Milberg et al. 2016, Seibold et al. 2016, Selberg 2019, Procházka and Schlaghamerský 2019, Parisi et al. 2021, Vogel et al. 2021, Mazur et al. 2021, Graf et al. 2022). Out of the 198 beetle species identified from birch dead wood, about a half (91 species) were also presented in beetle species lists obtained from Germany (Vogel et al. 2021), Poland (Sawoniewicz 2013), Norway (Kaila 1993, Vindstad et al. 2014) and Sweden (Lindhe and Lindelöw 2004). Amongst 210 beetle species collected from common oak deadwood, a total of 116 overlapping with beetle species lists were obtained in oaks in Germany (Vogel et al. 2021), France (Bouget et al. 2012, Lassauce et al. 2013) and Sweden (Lindhe and Lindelöw 2004, Milberg et al. 2016). Fifty-five beetle species collected from dead aspen overlapped when compared with research conducted in Germany (Vogel et al. 2021), Finland (Martikainen 2001, Ranius et al. 2011) and Sweden (Lindhe and Lindelöw 2004, Selberg 2019). We did not find many listings of beetles in dead wood of small-leaved linden. However, 48 beetle species collected in our research overlap with species obtained in several works (Jonsell 2012, Vogel et al. 2021). Limited research and species lists make it difficult to compare our findings on beetles in dead wood of European ashes and black alders. However, comparing our study with research in Germany, we found overlapping beetle species in both trees: three in European ashes and four in black alders (Vogel et al. 2021). Although the number of overlapping species is relatively small, this suggests that the beetle species in dead European ash and black alder may be much more similar than they appear to be at present, but further research on the dead wood of these tree species is needed. The comparison of beetle lists and other researchers' findings highlights the significance of tree species of beetle communities (Zuo et al. 2021).

Based on other authors' species lists and species biology, beetle species regarded as saproxylic (according to Speight (1989)) accounted for about a half of our collected beetles, while the remaining are considered as non-wood dependent species (Hågvar and Økland 1997, Schmidl and Bußler 2004, Wikars et al. 2005, Byk et al. 2006, Gibb et al. 2006, Brunet and Isacsson 2008, Mařák and Schlaghamerský 2009, Horák 2011, De Biase 2011, Vodka and Cizek 2013, Sawoniewicz 2013, Milberg et al. 2014, Vindstad et al. 2014, Anonymous 2015, Papis and Mokrzycki 2015, Carlsson et al. 2016, Seibold et al. 2016, Procházka and Schlaghamerský 2019, Marker 2019, Ekström 2020). Our research, as well as that of other scientists, includes a range of beetle species whose biology is not directly related to this unique habitat and they can be considered incidental to dead wood. Research from neighbouring Poland reveals (Anonymous 2015) that the most of the non-wood-dependent beetles we have collected are associated with accumulated organic matter, plant litter, mud and soil, various plants, mosses, the nests of birds and other animals, carrion and fungal fruiting bodies (Table 2). Staphylinidae was one of the most diverse families in our research. About half of the rove beetles collected are not associated with dead wood and were characterised by relatively low abundance (Table 2).

The high number of non-saproxylic as well as saproxylic beetle species that we found associated with dead deciduous tree species in Lithuania highlights the importance of dead wood for conservation of the overall forest beetle community.

## Conflicts of interest

The authors have declared that no competing interests exist.

## References

- Abrahamsson M, Jonsell M, Niklasson M, Lindbladh M (2009) Saproxylic beetle assemblages in artificially created high-stumps of spruce (*Picea abies*) and birch (*Betula pendula/pubescens*) - does the surrounding landscape matter? Insect Conservation and Diversity 2 (4): 284-294. <https://doi.org/10.1111/j.1752-4598.2009.00066.x>
- Anonymous (2015) Coleoptera Poloniae, KFP base. URL: <https://coleoptera.ksib.pl/kfp.php?l=en>
- Bakke A (1999) High diversity of saproxylic beetles in a hemiboreal mixed forest reserve in the south of Norway. Scandinavian Journal of Forest Research 14 (3): 199-208. <https://doi.org/10.1080/02827589950152719>
- Bouget C, Brustel H, Brin A, Noblecourt T (2008) Sampling saproxylic beetles with window flight traps: methodological insights. Revue d'Écologie (La Terre et La Vie) 10 (1): 21-32. <https://doi.org/10.3406/revec.2008.1457>
- Bouget C, Nusillard B, Pineau X, Ricou C (2012) Effect of deadwood position on saproxylic beetles in temperate forests and conservation interest of oak snags: Oak snags and saproxylic beetle conservation. Insect Conservation and Diversity 5 (4): 264-278. <https://doi.org/10.1111/j.1752-4598.2011.00160.x>
- Brunet J, Isacsson G (2008) Influence of snag characteristics on saproxylic beetle assemblages in a south Swedish beech forest. Journal of Insect Conservation 13 (5): 515-528. <https://doi.org/10.1007/s10841-008-9200-3>
- Byk A, Mokrzycki T, Perliński S, Rutkiewicz A (2006) Saproxylic beetles – in the monitoring of anthropogenic transformations of Białowieża primeval forest. Warsaw Agricultural University Press
- Carlsson S, Bergman K, Jansson N, Ranius T, Milberg P (2016) Boxing for biodiversity: evaluation of an artificially created decaying wood habitat. Biodiversity and Conservation 25 (2): 393-405. <https://doi.org/10.1007/s10531-016-1057-2>
- De Biase A (2011) First contribution to the knowledge of the shining beetles of Sardinia (Coleoptera: Phalacridae). Conservazione Habitat Invertebrati 5: 493-500.
- De Jong Y, Verbeek M, Michelsen V, Bjørn PDP, Los W, Steeman F, Bailly N, Basire C, Chylarecki P, Stloukal E, Hagedorn G, Wetzel F, Glöckler F, Kroupa A, Korb G, Hoffmann A, Häuser C, Kohlbecker A, Müller A, Güntsch A, Stoev P, Penev L (2014) Fauna Europaea – all European animal species on the web. Biodiversity Data Journal 2 <https://doi.org/10.3897/BDJ.2.e4034>

- Djupström L, Weslien J, Schroeder LM (2008) Dead wood and saproxylic beetles in set-aside and non set-aside forests in a boreal region. *Forest Ecology and Management* 255 (8-9): 3340-3350. <https://doi.org/10.1016/j.foreco.2008.02.015>
- Edelmann P, Ambarlı D, Gossner M, Schall P, Ammer C, Wende B, Schulze E, Weisser W, Seibold S (2022) Forest management affects saproxylic beetles through tree species composition and canopy cover. *Forest Ecology and Management* 524 <https://doi.org/10.1016/j.foreco.2022.120532>
- Ekström LA (2020) The importance of Ecoparks for saproxylic beetles: a study on general ecological hypotheses in differently managed landscapes. Swedish University of Agricultural Sciences, SLU Department of Wildlife, Fish, and Environmental Studies, Sweden.
- Ferenca R, Tamutis V (2011) Contribution to the knowledge of Lithuanian Silvanidae Kirby, 1837 (Insecta: Coleoptera). New and Rare for Lithuania Insect Species 23: 23-29.
- Ferenca R, Ivinskis P, Meržijevskis A, Rimšaitė J, Karalius S (2011) Twenty beetle (Insecta: Coleoptera) species new for the Lithuanian fauna. New and Rare for Lithuania Insect Species 23: 15-22.
- Ferenca R, Tamutis V, Kinduris R (2013) New records of rare false blister beetle (Coleoptera: Oedemeridae) species in Lithuania. New and Rare for Lithuania Insect Species 25: 10-18.
- Ferenca R, Tamutis V (2015) New data on *Agabus* species (Coleoptera: Dytiscidae) in Lithuania. New and Rare for Lithuania Insect Species 27: 18-23.
- Ferenca R, Tamutis V, Inokaitis V, Martinaitis K (2016) Data on beetle (Coleoptera) species new to Lithuanian fauna. New and Rare for Lithuania Insect Species 28: 21-31.
- Ferenca R, Tamutis V, Inokaitis V, Martinaitis K (2018) Data on new and insufficiently known for Lithuanian fauna species of beetles (Insecta: Coleoptera). Bulletin of the Lithuanian Entomological Society 2: 16-25.
- Gibb H, Pettersson RB, Hjältén J, Hilszczański J, Ball JP, Johansson T, Atlegrim O, Danell K (2006) Conservation-oriented forestry and early successional saproxylic beetles: Responses of functional groups to manipulated dead wood substrates. *Biological Conservation* 129 (4): 437-450. <https://doi.org/10.1016/j.biocon.2005.11.010>
- Government of the Republic of Lithuania (2022) National forestry accounting 2022.01.01. URL: <https://amvmt.lrv.lt/lv/atviri-duomenys-1/misku-statistikos-leidiniai/valstybine-misku-apskaita/2022-01-01>
- Graf M, Lettenmaier L, Müller J, Hagge J (2022) Saproxylic beetles trace deadwood and differentiate between deadwood niches before their arrival on potential hosts. *Insect Conservation and Diversity* 15 (1): 48-60. <https://doi.org/10.1111/icad.12534>
- Hågvar S, Økland B (1997) Saproxylic beetle fauna associated with living sporocarps of *Fomitopsis pinicola* (Fr.) Karst. in four spruce forests with different management histories. *Fauna Norvegica Series B* 44: 95-105.
- Halme P, Vartiainen N, Salmela J, Penttinen J, Norros V (2013) High within- and between-trunk variation in the nematoceran (Diptera) community and its physical environment in decaying aspen trunks. *Insect Conservation and Diversity* 6 (4): 502-512. <https://doi.org/10.1111/icad.12007>
- Hjältén J, Stenbacka F, Andersson J (2010) Saproxylic beetle assemblages on low stumps, high stumps and logs: Implications for environmental effects of stump harvesting. *Forest Ecology and Management* 260 (7): 1149-1155. <https://doi.org/10.1016/j.foreco.2010.07.003>

- Horák J (2011) Response of saproxylic beetles to tree species composition in a secondary urban forest area. *Urban Forestry & Urban Greening* 10 (3): 213-222. <https://doi.org/10.1016/j.ufug.2011.04.002>
- Irmler U, Heller K, Warning J (1996) Age and tree species as factors influencing the populations of insects living in dead wood (Coleoptera, Diptera: Sciaridae, Mycetophilidae). *Pedobiologia* 40: 134-148.
- Ivinskis P, Rimšaitė J, Meržijevskij A (2013) Data on beetle (Coleoptera) species new for Lithuanian fauna. *New and Rare for Lithuania Insect Species* 25: 18-23.
- Ivinskis P, Meržijevskij A, Rimšaitė J (2014) Data about new beetle (Coleoptera) species found in Lithuania. *New and Rare for Lithuania Insect Species* 26: 31-36.
- Ivinskis P, Rimšaitė J, Meržijevskij A (2015) New species and new records of rare species of beetles (Coleoptera) from Lithuania. *New and Rare for Lithuania Insect Species* 27: 27-34.
- Ivinskis P, Rimšaitė J, Meržijevskij A, Jefanovas A (2017) New records of new and rare beetle (Coleoptera) species for Lithuania. *Bulletin of the Lithuanian Entomological Society* 1: 17-22.
- Jacobsen RM, Birkemoe T, Sverdrup-Thygeson A (2015) Priority effects of early successional insects influence late successional fungi in dead wood. *Ecology and Evolution* 5 (21): 4896-4905. <https://doi.org/10.1002/ece3.1751>
- Jonsell M, Weslien J, Ehnström B (1998) Substrate requirements of red-listed saproxylic invertebrates in Sweden. *Biodiversity and Conservation* 7 (6): 749-764. <https://doi.org/10.1023/A:1008888319031>
- Jonsell M, Nittérus K, Stighäll K (2004) Saproxylic beetles in natural and man-made deciduous high stumps retained for conservation. *Biological Conservation* 118 (2): 163-173. <https://doi.org/10.1016/j.biocon.2003.08.017>
- Jonsell M, Hansson J (2007) Comparison of methods for sampling saproxylic beetles in fine wood. *Entomologica Fennica* 18 (4): 232-241. <https://doi.org/10.33338/ef.84404>
- Jonsell M (2012) Old park trees as habitat for saproxylic beetle species. *Biodiversity and Conservation* 21 (3): 619-642. <https://doi.org/10.1007/s10531-011-0203-0>
- Jonsson B, Kruys N, Ranius T (2005) Ecology of species living on dead wood – lessons for dead wood management. *Silva Fennica* 39 (2): 289-309. <https://doi.org/10.14214/sf.390>
- Kaila L (1993) A new method for collecting quantitative samples of insects associated with decaying wood or wood fungi. *Entomologica Fennica* 4 (1): 21-23. <https://doi.org/10.33338/ef.83745>
- Lassauze A, Larrieu L, Paillet Y, Lieutier F, Bouget C (2013) The effects of forest age on saproxylic beetle biodiversity: implications of shortened and extended rotation lengths in a French oak high forest. *Insect Conservation and Diversity* 6 (3): 396-410. <https://doi.org/10.1111/j.1752-4598.2012.00214.x>
- Lekoveckaitė A, Podienė V, Ferenca R (2017) New records of *Phyllodrepa* (*Phyllodrepa*) melanocephala Fabricius, 1787 and *Telmatophilus brevicollis* Aubé, 1862 (Coleoptera: Staphylinidae and Cryptophagidae) in Lithuania. *Bulletin of the Lithuanian Entomological Society* 1: 23-26.
- Lekoveckaitė A, Podienė V, Ferenca R (2019) New and rare for Lithuanian fauna beetle species found in Büda botanical – zoological and Biržai forest botanical reserves, report of 2018. *Bulletin of the Lithuanian Entomological society* 3: 19-24.

- Lindhe A, Lindelöw Å (2004) Cut high stumps of spruce, birch, aspen and oak as breeding substrates for saproxylic beetles. Forest Ecology and Management 203 (1-3): 1-20. <https://doi.org/10.1016/j.foreco.2004.07.047>
- Macagno AM, Hardersen S, Nardi G, Lo Giudice G, Mason F (2015) Measuring saproxylic beetle diversity in small and medium diameter dead wood: The "grab-and-go" method. European Journal of Entomology 112 (3): 510-519. <https://doi.org/10.14411/eje.2015.049>
- Mařák V, Schlaghamerský J (2009) The saproxylic beetles of Dlúhý hrúd, an old-growth remnant on the Lower Dyje River (Czechia). In: Buse J, Alexander KNA, Ranius T, Assmann T (Eds) Saproxylic beetles - their role and diversity in European woodland and tree habitats. Pensoft Publishers, Sofia.
- Marker JD (2019) Effect of distance to urban areas on saproxylic beetles in urban forests. Karlstad University, Faculty of Social and Life Sciences, Department of Biology, Sweden.
- Martikainen P (2001) Conservation of threatened saproxylic beetles: Significance of retained aspen *Populus tremula* on clearcut areas. Ecological Bulletins 49: 205-218.
- Mazur A, Witkowski R, Kuźmiński R, Jaszczałk R, Turski M, Kwaśna H, Łakomy P, Szmyt J, Adamowicz K, Łabędzki A (2021) The structure of saproxylic beetle assemblages in view of coarse woody debris resources in pine stands of western Poland. Forests 12 (11): 1558. <https://doi.org/10.3390/f12111558>
- Milberg P, Bergman K, Johansson H, Jansson N (2014) Low host-tree preferences among saproxylic beetles: a comparison of four deciduous species. Insect Conservation and Diversity 7 (6): 508-522. <https://doi.org/10.1111/icad.12074>
- Milberg P, Bergman K, Sancak K, Jansson N (2016) Assemblages of saproxylic beetles on large downed trunks of oak. Ecology and Evolution 6 (6): 1614-1625. <https://doi.org/10.1002/ece3.1935>
- Monsevičius V (2013) New and little known for the Lithuanian fauna species of beetles (Coleoptera), found in 2002, 2011–2012. New and Rare for Lithuania Insect Species 25: 24-30.
- Monsevičius V (2020) Species of Staphylinidae (Coleoptera) new to the fauna of Lithuania. Bulletin of the Lithuanian Entomological Society 4: 9-19.
- Monsevičius V (2022) Twelve rove beetle (Coleoptera, Staphylinidae) species new to the Lithuanian fauna, found in 2020–2021. Bulletin of the Lithuanian Entomological Society 5: 22-28.
- Müller J, Hothorn T, Pretzsch H (2007) Long-term effects of logging intensity on structures, birds, saproxylic beetles and wood-inhabiting fungi in stands of European beech *Fagus sylvatica* L. Forest Ecology and Management 242 (2-3): 297-305. <https://doi.org/10.1016/j.foreco.2007.01.046>
- Müller J, Bussler H (2008) Key factors and critical thresholds at stand scale for saproxylic beetles in a beech dominated forest, southern Germany. Revue d'Écologie (La Terre et La Vie) 10 (1): 81-90. <https://doi.org/10.3406/revec.2008.1464>
- Müller J, Wende B, Strobl C, Eugster M, Gallenberger I, Floren A, Steffan-Dewenter I, Linsenmair KE, Weisser W, Gossner M (2015) Forest management and regional tree composition drive the host preference of saproxylic beetle communities. Journal of Applied Ecology 52 (3): 753-762. <https://doi.org/10.1111/1365-2664.12421>
- Müller J, Ulyshen M, Seibold S, Cadotte M, Chao A, Bässler C, Vogel S, Hagge J, Weiß I, Baldrian P, Tláskal V, Thorn S (2020) Primary determinants of communities in

- deadwood vary among taxa but are regionally consistent. *Oikos* 129 (10): 1579-1588. <https://doi.org/10.1111/oik.07335>
- Muñoz-López NZ, Andrés-Hernández AR, Carrillo-Ruiz H, Rivas-Arancibia SP (2016) Coleoptera associated with decaying wood in a tropical deciduous forest. *Neotropical Entomology* 45 (4): 341-350. <https://doi.org/10.1007/s13744-016-0367-0>
  - Nagrockaitė R, Tamutė B, Tamutis V (2011) New and rare beetle (Coleoptera) species from Curonian spit (Lithuania). *New and Rare for Lithuania Insect Species* 23: 34-38.
  - Pacevičius V (2017) New for Lithuanian fauna species of beetles (Coleoptera) found in Molėtai district. *Bulletin of the Lithuanian Entomological Society* 1: 27-29.
  - Paletto A, Tosi V (2010) Deadwood density variation with decay class in seven tree species of the Italian Alps. *Scandinavian Journal of Forest Research* 25 (2): 164-173. <https://doi.org/10.1080/02827581003730773>
  - Palviainen M, Laiho R, Mäkinen H, Finér L (2008) Do decomposing Scots pine, Norway spruce, and silver birch stems retain nitrogen? *Canadian Journal of Forest Research* 38 (12): 3047-3055. <https://doi.org/10.1139/X08-147>
  - Papis M, Mokrzycki T (2015) Saproxylic beetles (Coleoptera) of the strictly protected area Bukowa Góra in the Roztoczański National Park. *Forest Research Papers* 76 (3): 229-239. <https://doi.org/10.48538/FRP-2015-0022>
  - Parisi F, Innangi M, Tognetti R, Lombardi F, Chirici G, Marchetti M (2021) Forest stand structure and coarse woody debris determine the biodiversity of beetle communities in Mediterranean mountain beech forests. *Global Ecology and Conservation* 28: e01637. <https://doi.org/10.1016/j.gecco.2021.e01637>
  - Paukkunen J, Biström O, Budrys E, Helve E, Lagercrantz CG, Mannerkoski I, Orlovskytė S, Tähtinen M (2016) Entomological excursion to the Curonina spit in august 2016. *New and Rare for Lithuania Insect Species* 28: 97-120.
  - Peuhu E, Thomssen P, Siitonens J (2019) Comparison of three trap types in sampling saproxylic beetles living in hollow urban trees. *Journal of Insect Conservation* 23 (1): 75-87. <https://doi.org/10.1007/s10841-018-0115-3>
  - Procházka J, Schlaghamerský J (2019) Does dead wood volume affect saproxylic beetles in montane beech-fir forests of Central Europe? *Journal of Insect Conservation* 23 (1): 157-173. <https://doi.org/10.1007/s10841-019-00130-4>
  - Ranius T, Jansson N (2002) A comparison of three methods to survey saproxylic beetles in hollow oaks. *Biodiversity and Conservation* 11 (10): 1759-1771. <https://doi.org/10.1023/A:1020343030085>
  - Ranius T, Martikainen P, Kouki J (2011) Colonisation of ephemeral forest habitats by specialised species: beetles and bugs associated with recently dead aspen wood. *Biodiversity and Conservation* 20 (13): 2903-2915. <https://doi.org/10.1007/s10531-011-0124-y>
  - Redolfi De Zan L, Bellotti F, D'Amato D, Carpaneto GM (2014) Saproxylic beetles in three relict beech forests of central Italy: Analysis of environmental parameters and implications for forest management. *Forest Ecology and Management* 328: 229-244. <https://doi.org/10.1016/j.foreco.2014.05.040>
  - Renvall P (1995) Community structure and dynamics of wood-rotting Basidiomycetes on decomposing conifer trunks in northern Finland. *Karstenia* 35 (1): 1-51. <https://doi.org/10.29203/ka.1995.309>

- Sawoniewicz M (2013) Beetles (Coleoptera) occurring in decaying birch (*Betula* spp.) wood in the Kampinos National Park. Forest Research Papers 74 (1): 71-85. <https://doi.org/10.2478/frp-2013-0008>
- Schmidl J, Bußler H (2004) Ecological guilds of xylobiontic (saproxyllic) beetles in Germany and their use in landscape ecological surveys - A methodical standard. Naturschutz und Landschaftsplanung 36 (7): 202-217.
- Seibold S, Bässler C, Brandl R, Büche B, Szallies A, Thorn S, Ulyshen M, Müller J (2016) Microclimate and habitat heterogeneity as the major drivers of beetle diversity in dead wood. Journal of Applied Ecology 53 (3): 934-943. <https://doi.org/10.1111/1365-2664.12607>
- Selberg S (2019) Saproxylic beetles in *Populus tremula* fauna depots – how do you construct the best depot? Biology Education Centre, Uppsala University and Sveriges Lantbruksuniversitet, Uppsala Kommun, Sweden.
- Speight MC (1989) Saproxylic invertebrates and their conservation. Council of Europe, France, Strasbourg.
- Stokland JN, Siitonens J, Jonsson BG (2012) Biodiversity in dead wood. Cambridge University Press, New York. <https://doi.org/10.1017/CBO9781139025843>
- Sverdrup-Thygeson A, Skarpaas O, Ødegaard F (2010) Hollow oaks and beetle conservation: the significance of the surroundings. Biodiversity and Conservation 19 (3): 837-852. <https://doi.org/10.1007/s10531-009-9739-7>
- Tamutis V, Tamutė B, Ferencia R (2011) A catalogue of Lithuanian beetles (Insecta: Coleoptera). ZooKeys 121: 1-494. <https://doi.org/10.3897/zookeys.121.732>
- Tamutis V (2012) New and rare (insufficiently known) beetle species found in the litter of coniferous and mixed forests in Lithuania. New and Rare for Lithuania Insect Species 24: 7-17.
- Tamutis V, Barševskis A (2014) A faunistic review of ground beetles of Lebiinae Bonelli, 1810 (Coleoptera: Carabidae) of Lithuania. Entomologica Fennica 25: 66-86.
- Tamutis V, Tamutė B, Ferencia R (2015) New data on rare and insufficiently known species of ground beetles (Coleoptera: Carabidae: Nebriinae, Carabinae, Omophroninae, Elaphrininae, Scaritinae, Broscinae) in Lithuania. New and Rare for Lithuania Insect Species 27: 47-62.
- Tamutis V, Martinaitis K (2019) *Grammoptera abdominalis* (Stephens, 1831) (Coleoptera: Cerambycidae) – longhorn beetle species new for Lithuanian fauna. Bulletin of the Lithuanian Entomological Society 3: 38-41.
- Toivanen T, Kotiaho J (2010) The preferences of saproxyllic beetle species for different dead wood types created in forest restoration treatments. Canadian Journal of Forest Research 40 (3): 445-464. <https://doi.org/10.1139/X09-205>
- Unal S, Ozcan E, Kaygin AT (2009) Wood-destroying Coleopteran species in the historical buildings in Kastamonu, in Turkey. African Journal of Biotechnology 8: 2349-2355.
- Varnagirytė-Kabašinskienė I, Lukminė D, Mizaras S, Beniušienė L, Armolaitis K (2019) Lithuanian forest biomass resources: legal, economic and ecological aspects of their use and potential. Energy, Sustainability and Society 9 (1): 1-19. <https://doi.org/10.1186/s13705-019-0229-9>
- Vindstad OPL, Schultze S, Jepsen JU, Biuw M, Kapari L, Sverdrup-Thygeson A, Ims RA (2014) Numerical responses of saproxyllic beetles to rapid increases in dead wood

- availability following geometrid moth outbreaks in sub-arctic mountain birch forest. PLoS One 9 (6): e99624. <https://doi.org/10.1371/journal.pone.0099624>
- Vodka Š, Cizek L (2013) The effects of edge-interior and understorey-canopy gradients on the distribution of saproxylic beetles in a temperate lowland forest. Forest Ecology and Management 304: 33-41. <https://doi.org/10.1016/j.foreco.2013.04.007>
  - Vogel S, Bussler H, Finnberg S, Müller J, Stengel E, Thorn S (2021) Diversity and conservation of saproxylic beetles in 42 European tree species: an experimental approach using early successional stages of branches. Insect Conservation and Diversity 14 (1): 132-143. <https://doi.org/10.1111/icad.12442>
  - Wende B, Gossner M, Grass I, Arnstadt T, Hofrichter M, Floren A, Linsenmair KE, Weisser W, Steffan-Dewenter I (2017) Trophic level, successional age and trait matching determine specialization of deadwood-based interaction networks of saproxylic beetles. Proceedings of the Royal Society B: Biological Sciences 284 (1854): 20170198. <https://doi.org/10.1098/rspb.2017.0198>
  - Wikars L, Sahlin E, Ranius T (2005) A comparison of three methods to estimate species richness of saproxylic beetles (Coleoptera) in logs and high stumps of Norway spruce. The Canadian Entomologist 137 (3): 304-324. <https://doi.org/10.4039/n04-104>
  - Zuo J, Berg M, Van Hal J, Van Logtestijn RP, Goudzwaard L, Hefting M, Poorter L, Sterck F, Cornelissen JC (2021) Fauna community convergence during decomposition of deadwood across tree species and forests. Ecosystems 24 (4): 926-938. <https://doi.org/10.1007/s10021-020-00558-9>