

Research Article

The first complete mitochondrial genome in the family Attevidae (*Atteva aurea*) of the order Lepidoptera

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Abstract

The superfamily Yponomeutoidea, one of the early-derived groups in the order Lepidoptera, consists of 11 families. However, mitochondrial genome (mitogenome) sequences, popularly used for phylogeny and evolutionary tracing, are available for only seven species across six genera and five families. Thus, a larger variety of mitogenome sequences in Yponomeutoidea are required to improve our understanding of lepidopteran phylogeny and genomic evolution. In this study, we present the complete mitogenome of *Atteva aurea* (Fitch, 1856), the first species in the family Attevidae (superfamily Yponomeutoidea, order Lepidoptera) to be sequenced. The complete mitogenome comprises 16,329 bp and contains a typical set of genes and one non-coding region. Within Yponomeutoidea, the mitogenome of *A. aurea* has a unique *trnl-trnM-trnQ* arrangement at the A + T-rich region and *ND2* junction and *trnA-ND3* arrangement at the *trnG* and *trnR* junction. Twelve of the 13 protein-coding genes (PCGs) of *A. aurea* have a typical ATN starting codon, whereas *COI* has the atypical CGA codon, which is frequently found in the starting region of lepidopteran *COI*. Phylogenetic analyses, based on the

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concatenated sequences of 13 PCGs and two rRNA genes, using the Maximum Likelihood method, revealed a sister relationship between Attevidae and Praydidae with moderately low nodal support (bootstrap support = 64%).

Keywords

mitochondrial genome, Atteva aurea, phylogeny, Attevidae

Introduction

The superfamily Yponomeutoidea is one of the earliest groups to develop external feeding mechanisms in the order Lepidoptera and comprises ~ 1,800 species across 11 families (Sohn et al. 2013). However, only seven species in six genera across five families have available mitochondrial genome (mitogenome) sequences. Thus, the characterisation of the mitogenomes of more families will significantly contribute to the study of genomic evolution and subsequent phylogenetic analysis within this superfamily, as well as other early-derived lepidopteran clades.

The ailanthus webworm (*Atteva aurea* Fitch, 1856) is a small, colourful moth predominantly found north of Costa Rica, across the USA and in southern Quebec and Ontario, Canada (Wilson et al. 2010). Populations distributed south of Costa Rica in Uruguay and Argentina are known as *A. pustulella* (Fabricius, 1787), the former classification of *A. aurea* in North America (Wilson et al. 2010).

In this study, we present the complete mitogenome of *A. aurea*, the first species in the family Attevidae (superfamily Yponomeutoidea, order Lepidoptera) to be sequenced. The sequence was analysed in terms of its mitogenome characteristics and phylogenetic position within the superfamily Yponomeutoidea. Additionally, the DNA barcoding region of *A. aurea* was compared to that of previously-registered *A. aurea* and *A. pustulella*, which have been used for extensive phylogenetic analysis (Wilson et al. 2010), to further confirm sequence divergence between the two species.

Materials and methods

Sample collection, DNA extraction, PCR and sequencing

In 2011, a brood of *A. aurea* was collected from the Paint Branch Trail at the University of Maryland (College Park, MA, USA; 38°59'39"N, 76°56'5"W). In this study, DNA was extracted from the whole body of one adult male using the Wizard Genomic DNA Purification Kit (Promega, Madison, WI, USA). Using Lepidoptera-specific primers (Suppl. material 1, Kim et al. 2012), three overlapping long fragments (LFs; *COI* to *ND4*, *ND5* to *IrRNA* and *IrRNA* to *COI*) were amplified. These LFs were then used as templates for the amplification of 26 short fragments (SFs) using the same Lepidoptera-specific primers (Suppl. material 1, Kim et al. 2012). All products were sequenced in both forward and

reverse transcriptional directions by Sanger's methods. The whole body of the specimen was consumed in the process. Thus, other individuals of the brood were moved as voucher specimens to the Gongju National University of Education (Gongju, South Korea) and labelled with accession nos. GNUE-I-0001–GNUE-I-0003.

Boundary delimitation and annotation

Individual SF sequences were manually assembled into complete mitogenomes using SeqMan (DNASTAR, Madison, WI, USA). The identification and boundary delimitation of each gene and secondary structure folding of tRNAs were performed using the MITOS Web Server (http://mitos.bioinf.uni-leipzig.de/index.py) and using the default search mode, Mito/Chloroplast as the searching source and the genetic code of invertebrate mitogenomes for tRNA isotype prediction (Lowe and Chan 2016). Where necessary, mitogenome sequences of species in the superfamily Yponomeutoidea registered in GenBank were downloaded and aligned for improved annotation by following the protocols presented by Cameron 2014.

Phylogenetic analysis

Phylogenetic analysis was conducted using 25 available mitogenomes in 23 species (including *A. aurea*) in the superfamilies Gracillarioidea, Yponomeutoidea and Tineoidea. We selected Gracillarioidea and Tineoidea, along with Yponomeutoidea, because of the previously established sister-group relationship between Yponomeutoidea and Gracillarioidea and of the branching of Tineoidea as a lineage basal to these two superfamilies (Timmermans et al. 2014, Breinholt et al. 2018, Bao et al. 2019, *Kawahara et al. 2019*). Two species within the superfamily Nepticuloidea (*Stigmella roborella* and *Astrotischeria* sp.) were used as outgroups. Thirteen protein-coding genes (PCGs) and two rRNA genes (including those of two outgroup species) were aligned using RevTrans ver. 2.0 (Wernersson and Pedersen 2003) and concatenated using SequenceMatrix ver. 1.8 (Vaidya et al. 2011). The Maximum Likelihood method was applied using CIPRES Portal ver. 3.1 (Miller et al. 2010) for phylogenetic analyses, based on the GTR + Gamma + I model, which was selected using jModelTest (Posada 2008).

Data resources

Genome sequence data used in this study are openly available from the GenBank database of the National Center for Biotechnology Information (<u>https://www.ncbi.nlm.nih.gov</u>) under the accession no. ON480203. All datasets used in this study were published at Zenodo on 03 July 2022 (Zenodo. <u>https://doi.org/10.5281/zenodo.6791899</u>).

Results and discussion

Genome summary

The complete 16,392-bp mitogenome of *A. aurea* is composed of typical gene sets (two rRNAs, 22 tRNAs and 13 PCGs) and a major non-coding A + T-rich region (Table 1; GenBank accession no. ON480203). Twelve of the 13 PCGs have the typical ATN start codon, whereas *COI* has the atypical CGA codon, which is found in the majority of other available mitogenome sequences of Tineoidea, Gracillarioidea and Yponomeutoidea species (data not shown), as well as Lepidoptera species (Kim et al. 2010, Park et al. 2016, Kim et al. 2018, Jeong et al. 2021). Ten PCGs end with typical stop codon TAA, whereas *COII* and *ND4* have incomplete, single-thymine stop codons. The A/T content is 79.9% in PCGs, 82.4% in tRNAs, 82.5% in the whole genome, 86.5% in *IrRNA*, 87.6% in *srRNA* and 98.0% in the A + T-rich region (data not shown).

Table 1.						
Summar	ry of <i>Atteva aurea</i> mito	chondrial	genome.			
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Gene	Nucleotide number	Size	Anticodon	Start codon	Stop codon	O/S
trnl	1-64	64	GAT 29-31			
trnM	65-132	68	CAT 96-98			-8
trnQ	141-209	69	TTG 177-179			-41
ND2	251-1276	1026		ATT	TAA	+2
trnW	1275-1340	64	TCA 1305-1307			+8
trnC	1333-1401	69	GCA 1369-1371			-7
trn Y	1409-1474	66	GTA 1441-1443			-5
COI	1480-3015	1536		CGA	TAA	-54
trnL ₂	3070-3136	67	TAA 3100-3102			
COII	3137-3818	682		ATG	T-tRNA	
trnK	3819-3889	71	CTT 3849-3851			+1
trnD	3889-3954	66	GTC 3919-3921			
ATP8	3955-4116	162		ATC	TAA	+7
ATP6	4110-4781	672		ATG	TAA	+1
COIII	4781-5569	789		ATG	TAA	-2
trnG	5572-5640	69	TCC 5602-5604			-332
trnA	5973-6039	67	TGC 6002-6004			-103
ND3	6143-6517	375		ATA	TAA	-28
trnR	6546-6607	62	TCG 6573-6575			-4
trnN	6612-6677	66	GTT 6642-6644			+1
trnS₁	6677-6737	61	GCT 6698-6700			-23

Gene	Nucleotide number	Size	Anticodon	Start codon	Stop codon	O/S
tmE	6761-6826	66	TTC 6791-6793			+2
tmF	6825-6892	68	GAA 6856-6858			-3
ND5	6896-8629	1716		ATT	TAA	
tmH	8630-8694	65	GTG 8659-8661			
ND4	8695-10036	1342		ATG	T-tRNA	+1
ND4L	10036-10323	288		ATG	TAA	-2
trnT	10326-10390	65	TGT 10357-10359			
tmP	10391-10455	65	TGG 10424-10426			-1
ND6	10457-10981	525		ATT	TAA	-3
CytB	10985-12136	1152		ATG	TAA	-10
trnS ₂	12147-12215	69	TGA 12179-12181			+2
ND1	12214-13170	957		ATG	TAA	-1
trnL1	13172-13238	67	TAG 13207-13209			
IrRNA	13239-14559	1321				
tmV	14560-14625	66	TAC 14594-14596			
srRNA	14626-15393	768				
A + T–rich region	15394-16392	999				

Non-underlined and underlined genes indicate forward and reverse transcriptional directions, respectively. tRNAs are denoted as one-letter symbols in accordance with the IUPAC-IUB single-letter amino acid codes, except those encoding leucine and serine, which are labelled L_1 for the CTN, L_2 for the TTR, S_1 for the AGN and S_2 for the TCN codon families. O/S denotes the number of the overlapping(+)/intergenic space sequence(-).

The genes of *A. aurea* are interleaved with a total 627 bp, spread over 17 regions ranging in size between 1 and 332 bp (Table 1). Most intergenic spacer sequences (ISSs) are short (1–20 bp), but four locations have longer ISSs (41–332 bp). Examination with the naked eye revealed that three of those four ISSs (*trnQ-ND2*, *COI- trnL*₂ and *trnA-ND3*) have no notable features, except high A/T content (90.24–96.30%; data not shown). However, the longest of these ISSs (332 bp), located between *trnG* and *trnA*, has four tandem repeat units with varying copy numbers (Fig. 1; A1–A4, B1–B3, C1–C12 and D1–D2). All copies of each repeat unit have identical sequences, except for one nucleotide substitution (A for G) in repeat unit D.



Four tandem repeat units found between tmG and tmA with various copy numbers (A1–A4, B1–B3, C1–C12 and D1–D2). The nucleotide position is indicated at each end of the sequence in relation to the mitochondrial genome of *Atteva aurea*.

Gene rearrangement

Compared with that of other Lepidoptera species, the A. aurea mitogenome has a very rare *trnl-trnM-trnQ* arrangement (underlining indicates gene inversion) at the A + T-rich region and ND2 junction (Fig. 2). Monopis longella (Walker, 1863) (family Tineidae, superfamily Tineoidea) is the only species previously known to exhibit the trnl-trnM-trnQ arrangement in the Ditrysia clade, including the superfamilies Gracillarioidea, Yponomeutoidea, and Tineoidea (Jeong et al. 2021). Conversely, the majority of ditrysian Lepidoptera species have the gene order trnM-trnI-trnQ at the same junction (Fig. 2, Kim et al. 2014). This differs from the ancestral *trnl-trnQ-trnM* order found in most insects (Fig. 2, Boore 1999), including ancient, non-ditrysian lepidopteran groups, such as Hepialoidea and Nepticuloidea (Cao et al. 2012, Timmermans et al. 2014). Moreover, the A. aurea mitogenome has the *trnA-ND3* arrangement at the *trnG* and *trnR* junctions instead of the ND3-trnA arrangement found in almost all Lepidoptera species, including all those in Gracillarioidea, Yponomeutoidea, and Tineoidea (Fig. 2, Kim et al. 2014, Jeong et al. 2021). Thus far, only seven species in six genera across fiv families in Yponomeutoidea have had their mitogenomes sequenced. Thus, further analysis of this superfamily is required to make any conclusive remarks about the evolution of this rearrangement. Nevertheless, current analysis indicates that the arrangement of the family Attevidae is an autapomorphic characteristic of the superfamily Yponomeutoidea (data not shown).

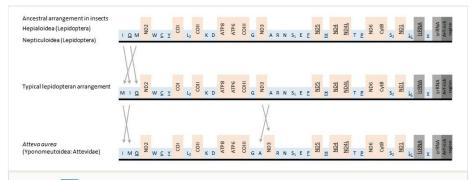


Figure 2. doi

Linear arrangement of the mitochondrial genome of *A. aurea*. Gene sizes are not drawn to scale. Non-underlined and underlined gene names indicate forward and reverse transcriptional directions, respectively. Translocated genes are indicated by lines with arrows.

Comparison of DNA barcoding sequence

The comparison between the DNA barcoding sequences of current *A. aurea* and those of *A. aurea* previously registered on GenBank, including those registered by Wilson et al. (2010), showed a 0.00–1.67% divergence. Compared with *A. pustulella* DNA barcoding sequences, there was a divergence of at least 3.95% (data not shown). This reflects the findings of a previous study that *A. aurea*, distributed between Costa Rica and southern Quebec and Ontario, is indeed *A. aurea* and that phylogenetic results demonstrate a clear

separation of *A. aurea* from other Attevidae species, including *A. pustulella* (Wilson et al. 2010).

Phylogenetic analysis

Phylogenetic analysis revealed overall lower nodal supports for familial relationships within Yponomeutoidea. A sister relationship between the families Attevidae and Praydidae, each of which is represented by a single species, was supported, but the nodal support for this relationship was not high (bootstrap support (BS) = 64%; Fig. 3). Within the Ditrysia clade, Gracillarioidea and Yponomeutoidea exhibit a sister relationship with the highest nodal support, placing Tineoidea sister to the two superfamilies with the highest support (Fig. 3). Previously, Sohn et al. (2013), using 8.0-18.9 kb of 8-27 genes from 11 families in Yponomeutoidea, also revealed a sister relationship between Attevidae and Praydidae and this relationship was supported with relatively high nodal support. In terms of relationships between superfamilies, our findings are consistent with previous studies based on mitogenomic, molecular, morphological, genomic and transcriptome data that proposed a sister-group relationship between Yponomeutoidea and Gracillarioidea, with Tineoidea diverging earlier within ditrysian Lepidoptera (Heikkilä et al. 2014, Timmermans et al. 2014, Breinholt et al. 2018, Bao et al. 2019, Kawahara et al. 2019). Additional phylogenetic relationships within the early-derived groups of Lepidoptera could be determined, based on further taxonomic research with a wider scope.

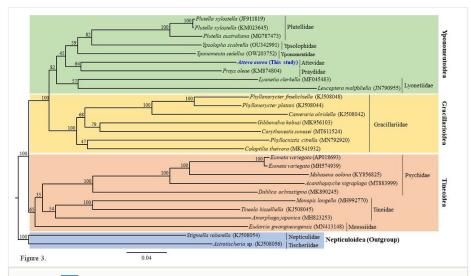


Figure 3. doi

Phylogenetic tree of the three ditrysian superfamilies included in this study: Tineoidea, Gracillarioidea and Yponomeutoidea. The number at each node indicates the bootstrap value. The scale bar indicates the number of substitutions per site. The GenBank accession number of each species is shown in brackets after its scientific name.

Conclusions

This mitogenome of *A. aurea* has a unique *trnl-trnM-trnQ* arrangement at the A + T-rich region and *ND2* junction and *trnA-ND3* arrangement at the *trnG* and *trnR* junction, which is unprecedented in Yponomeutoidea. Thus, additional mitogenome sequences are required from other genera, subfamilies and families to understand the taxonomic extent of this arrangement in Yponomeutoidea. Phylogenetic analysis revealed a sister relationship between Attevidae and Praydidae, consistent with the results of a previous large-scale molecular phylogenetic study, but nodal support was not high in this study. The result of our DNA barcoding sequence comparison supports the finding of a previous study that *A. aurea*, occurring north of Costa Rica in the USA and southern Quebec and Ontario, is genetically distinct from *A. pustulella* distributed from Costa Rica south to Uruguay and Argentina. Including that of *A. aurea*, only nine mitogenome sequences, representing seven genera across six families, are currently available for the superfamily Yponomeutoidea. Thus, more mitogenome sequences from the early-derived groups of Lepidoptera, including Yponomeutoidea, are essential for a greater understanding of mitogenome evolution and phylogenetic relationships in this order.

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Hosting institution

Chonnam National University, Gwangju, South Korea

Ethics and security

We declare that there are no violations of the guidelines of the authors' respective institutions and local, national and international regulations.

Author contributions

Conceptualisation, J.-C.S., J.S.J. and I.K.; fieldwork, J.-C.S.; data analysis and interpretation, J.S.J., J.S.P., H.K.O. and M.J.K.; writing—original draft, J.S.J., J.-C.S., M.J.K. and I.K.; writing—review and editing, J.-C.S., J.S.P., H.K.O. and I.K.; supervision, I.K.; project administration, J.S.P. and I.K.; funding acquisition, I.K.

Conflicts of interest

We declare no potential conflicts of interest.

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Supplementary material

Suppl. material 1: List of primers used to amplify and sequence the *Atteva aurea* mitochondrial genome doi

Authors: Jeong et al. Data type: Primer list Download file (33.10 kb)